

Review

# The Yin and Yang of I $\kappa$ B Kinases in Cancer

Abdalla M. Abdrabou <sup>1,2</sup>

<sup>1</sup> Department of Biochemistry and Molecular Genetics, Feinberg School of Medicine, Northwestern University, Chicago, IL 60611, USA; abdalla.abdrabou@northwestern.edu

<sup>2</sup> Department of Medical Genetics, University of Alberta, Edmonton, AB T6G 2H7, Canada

**Abstract:** I $\kappa$ B kinases (IKKs), specifically IKK $\alpha$  and IKK $\beta$ , have long been recognized for their pivotal role in the NF- $\kappa$ B pathway, orchestrating immune and inflammatory responses. However, recent years have unveiled their dual role in cancer, where they can act as both promoters and suppressors of tumorigenesis. In addition, the interplay with pathways such as the MAPK and PI3K pathways underscores the complexity of IKK regulation and its multifaceted role in both inflammation and cancer. By exploring the molecular underpinnings of these processes, we can better comprehend the complex interplay between IKKs, tumor development, immune responses, and the development of more effective therapeutics. Ultimately, this review explores the dual role of I $\kappa$ B kinases in cancer, focusing on the impact of phosphorylation events and crosstalk with other signaling pathways, shedding light on their intricate regulation and multifaceted functions in both inflammation and cancer.

**Keywords:** I $\kappa$ B kinases; IKKs; NF- $\kappa$ B



**Citation:** Abdrabou, A.M. The Yin and Yang of I $\kappa$ B Kinases in Cancer. *Kinases Phosphatases* **2024**, *2*, 9–27. <https://doi.org/10.3390/kinasesphosphatases2010002>

Academic Editors: Alison D. Axtman and Mauro Salvi

Received: 19 October 2023

Revised: 22 December 2023

Accepted: 29 December 2023

Published: 31 December 2023



**Copyright:** © 2023 by the author. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

## 1. Introduction

The NF- $\kappa$ B (Nuclear Factor- $\kappa$ B) pathway is a linchpin of cellular responses to external stimuli, especially in the realms of immune and inflammatory processes [1]. Within the broader context of the NF- $\kappa$ B signaling pathway, the I $\kappa$ B kinases (IKKs) (Table 1), particularly IKK $\alpha$  and IKK $\beta$ , occupy a pivotal position in the intricate regulatory network of these pathways. Traditionally recognized for their roles in immune surveillance and defense, the IKKs have recently emerged as enigmatic figures in the landscape of cancer biology [2–5]. Their Janus-faced nature, promoting or suppressing tumorigenesis depending on context, has prompted an intensive exploration of their molecular mechanisms within the context of cancer and the possibility of targeting them [6].

**Table 1.** Types of I $\kappa$ B kinases.

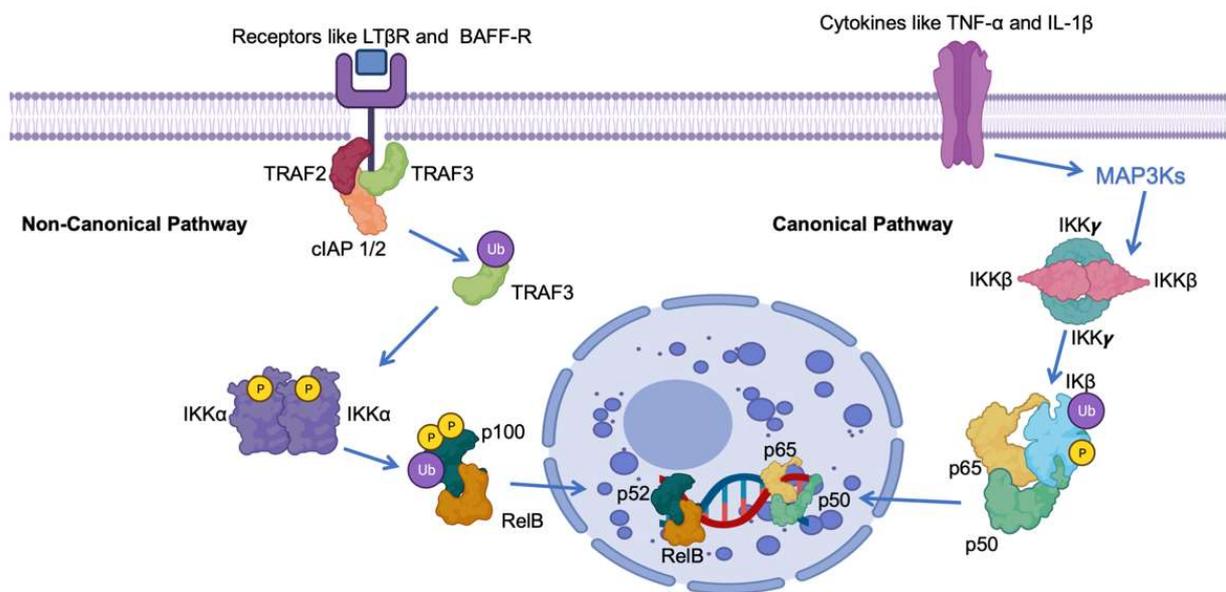
Type of I $\kappa$ B Kinase	Role in NF- $\kappa$ B Pathway	Function in NF- $\kappa$ B Regulation	Targets
IKK $\alpha$ (Inhibitor of $\kappa$ B Kinase Alpha) [7]	Non-Canonical NF- $\kappa$ B pathway	Phosphorylates p100, leading to partial proteasomal processing into p52. Initiates non-canonical gene transcription. Involved in cellular senescence.	p100
IKK $\beta$ (Inhibitor of $\kappa$ B Kinase Beta) [8]	Canonical NF- $\kappa$ B pathway	Phosphorylates I $\kappa$ B $\alpha$ and I $\kappa$ B $\beta$ , marking them for degradation. Releases p50-RelA dimers for nuclear translocation. Associated with chronic inflammation and tumor promotion.	I $\kappa$ B $\alpha$ , I $\kappa$ B $\beta$
IKK $\epsilon$ (Inhibitor of $\kappa$ B Kinase Epsilon) [9]	Both canonical and non-canonical pathways	Regulates NF- $\kappa$ B activation, particularly in response to viral infections. Can promote cell survival.	-
TBK1 (TANK-binding kinase 1) [10]	Non-Canonical NF- $\kappa$ B pathway	Activates IKK $\alpha$ and promotes non-canonical NF- $\kappa$ B signaling. Also involved in antiviral immune responses.	IKK $\alpha$
IKK $\zeta$ (Inhibitor of $\kappa$ B Kinase Zeta (MAIL)) [11]	Both canonical and non-canonical pathways	Modulates NF- $\kappa$ B signaling and immune responses. May play a role in inflammation and autoimmunity.	-
NEMO (NF- $\kappa$ B Essential Modulator) [12]	Central scaffold protein	Acts as an essential scaffold for IKK $\alpha$ and IKK $\beta$ , facilitating their activation. Essential for canonical NF- $\kappa$ B activation.	IKK $\alpha$ , IKK $\beta$

The NF- $\kappa$ B pathway, with its intricate family members, represents a dynamic signaling network that orchestrates cellular responses to a multitude of extracellular signals [13]. The fundamental aspect of this pathway revolves around the NF- $\kappa$ B transcription factors, comprising various members such as p65 (RelA), RelB, c-Rel, p105/p50, and p100/p52. These members of the NF- $\kappa$ B family combine in different dimeric forms, each playing specific roles in overseeing signaling pathways, particularly those involved in immune responses [14–16].

Central to the regulation of NF- $\kappa$ B is the presence of inhibitory proteins referred to as I $\kappa$ Bs, responsible for maintaining NF- $\kappa$ B dimers in an inactive state within the cell's cytoplasm [17]. The activation of the pathway involves the phosphorylation of I $\kappa$ Bs, tagging them for degradation by the proteasome machinery. This process liberates NF- $\kappa$ B dimers, allowing them to migrate into the nucleus and commence gene transcription (Figure 1) [18].

The NF- $\kappa$ B pathway, essential for various physiological processes, exists in multiple branches. Primarily, there are two well-characterized pathways: the canonical (or classical) and non-canonical (or alternative) [19]. These pathways differ in terms of the stimuli that activate them, the proteins involved, and the nature of their functions. The effective enhancement of NF- $\kappa$ B involves a complex regulation process mediated by the I $\kappa$ B kinase (IKK) complex. This intricate system orchestrates the phosphorylation of I $\kappa$ B proteins, leading to their ubiquitination and subsequent degradation via the proteasome [20–22]. This series of events ultimately results in the liberated NF- $\kappa$ B complexes translocating into the nucleus.

Within the nucleus, these NF- $\kappa$ B complexes engage with specific DNA sequences, thereby governing the transcription of genes involved in diverse processes, including immune responses, cellular growth regulation, and the modulation of cell survival [23–25]. Notably, within the context of cancer, NF- $\kappa$ B-dependent genes encompass those responsible for encoding cytokines, chemokines, cyclin D1, matrix metalloproteinases, and antiapoptotic proteins such as Bcl-xL [26–28].



**Figure 1.** Schematic representation of the roles of I $\kappa$ B in the canonical and non-canonical NF- $\kappa$ B pathway. The NF- $\kappa$ B pathway involves key proteins such as Tumor Necrosis Factor Receptor-Associated Factor 3 (TRAF3), Precursor protein 100 (p100), V-rel avian reticuloendotheliosis viral oncogene homolog B (RelB), Nuclear Factor NF-kappa-B p52 subunit (p52), Nuclear Factor NF-kappa-B p65 subunit (p65), Cellular Inhibitor of Apoptosis Protein 1/2 (cIAP 1/2), B-cell Activating Factor Receptor (BAFF-R), and Lymphotoxin Beta Receptor (LT $\beta$ R). Ub indicates ubiquitination, IKK  $\alpha$  (IkappaB kinase alpha). P indicates phosphorylation.

## 2. Canonical NF- $\kappa$ B Pathway

In the Canonical Nuclear Factor-kappa B (NF- $\kappa$ B) pathway, activation is initiated by a diverse array of stimuli, encompassing proinflammatory cytokines like Tumor Necrosis Factor-alpha (TNF- $\alpha$ ) and Interleukin-1 beta (IL-1 $\beta$ ), as well as microbial products like lipopolysaccharides (LPSs) (Figure 1) [29–32]. Key to this pathway is the activation of IKK $\beta$ , which subsequently phosphorylates and targets I $\kappa$ B $\alpha$  and I $\kappa$ B $\beta$  for degradation. As a result, this liberation allows for the movement of p50-RelA dimers into the nucleus. Once in the nucleus, these dimers function as transcription factors, overseeing the expression of genes linked to inflammation, immune responses, and cell survival (Figure 1) [33–35].

## 3. Non-Canonical NF- $\kappa$ B Pathway

Conversely, the Non-Canonical NF- $\kappa$ B pathway is typically triggered by a unique group of receptors, which encompass the lymphotoxin- $\beta$  receptor (LT $\beta$ R) and B-cell activating factor receptor (BAFF-R) [36]. This pathway is reliant on the conversion of p100 to p52, a process orchestrated by the activation of IKK $\alpha$ . Subsequently, the p52-RelB dimers relocate to the nucleus, assuming critical roles in the development of secondary lymphoid organs, B-cell maturation, and the organization of lymphoid tissues (Figure 1) [37–39].

NIK, or NF- $\kappa$ B-inducing kinase, holds a crucial position in regulating the non-canonical NF- $\kappa$ B signaling pathway [40]. Initially acknowledged for its role in activating the canonical NF- $\kappa$ B pathway, the absence of NIK did not hinder the TNF-induced IKK $\beta$ /p65/p50 activation. However, it was later discovered to be essential for triggering the non-canonical NF- $\kappa$ B pathway [41–43]. The regulation of NIK predominantly occurs post-translationally. Structurally, NIK encompasses four domains: a TRAF3-binding N-terminal region, a negative regulatory domain (NRD), a core kinase domain, and a C-terminal domain responsible for binding with proteins like IKK $\alpha$  and p100 [44–46].

Initially recognized as a mediator following TNF and IL-1 receptor activation, NIK's kinase activity was deemed crucial in facilitating this particular process. Additionally,

it mediates stimulation through various receptors like CD27, CD30, CD40, LT $\beta$ R, and BAFFR [47–50]. The overexpression of NIK activates NF- $\kappa$ B, protecting cells from TNF-induced apoptosis, while kinase-dead NIK mutants inhibit NF- $\kappa$ B activation by TNF $\alpha$  [51–53].

Under normal conditions, NIK binds to TRAF2/3 and cIAP1/2, leading to its continuous ubiquitination and degradation [54]. Stimulation by cytokines (such as CD40L, TWEAK, LT $\alpha$ / $\beta$ , or LPS) sequesters TRAF2/3, allowing the cIAP1-mediated ubiquitination of TRAF3 [55]. The subsequent degradation of TRAF3 leads to the accumulation of newly synthesized NIK within the cell. This stabilization and buildup of NIK are crucial for initiating the noncanonical NF- $\kappa$ B pathway [56–58]. Upon receptor activation, NIK triggers IKK $\alpha$  phosphorylation at Ser-176 and Ser-180, activating it to phosphorylate p100. The phosphorylation of p100 prompts the binding to ubiquitin ligase  $\beta$ -TrCP, resulting in partial proteasomal processing to p52. This processing removes the inhibitory C-terminal ankyrin repeat domain of p100, akin to the function of mature I $\kappa$ B proteins, thus maintaining RelB inactive in the cytoplasm. Subsequently, p52-RelB translocates to the nucleus to regulate transcription [59–62].

NIK interacts with and activates both IKK $\alpha$  and IKK $\beta$ , phosphorylating IKK $\alpha$  to a greater extent. Consequently, NIK acts as an upstream kinase for the IKK complex, facilitating signaling from multiple cytokine receptors [63–65]. Several other kinases, including MEKK1 and TAK1, were identified as IKK kinases, sometimes acting alongside NIK. TAK1, for instance, can activate NIK/IKK/NF- $\kappa$ B signaling independently of NIK in certain contexts [66]. Additionally, proteins like TRAF2, 5, and 6, as well as TBK-1, contribute to NF- $\kappa$ B activation by acting upstream of NIK. Moreover, Bcl10 has been reported to phosphorylate NIK under specific inflammatory conditions in human colonic epithelial cells treated with carrageenan (CGN) [22,67,68].

#### 4. The Dark Side: I $\kappa$ B Kinases as Tumor Promoters

##### 4.1. IKK $\alpha$ (Inhibitor of $\kappa$ B Kinase Alpha)

IKK $\alpha$  plays a multifaceted role in cancer, impacting both its initiation and progression, along with metastasis. In colorectal cancer cells (HT29), IKK $\alpha$  exhibits abnormal activation within the nucleus of tumor cells [69]. Here, it binds to specific genes reliant on Notch signaling, such as *hes1* and *herp2*. The nuclear IKK $\alpha$  phosphorylates a nuclear co-repressor, SMRT, causing its release from chromatin and the subsequent expression of Notch-dependent genes [70], leading to more aggressive growth and proliferation. Pan-IKK inhibition re-establishes SMRT chromatin binding, curbing Notch-related gene expression, and restraining tumor growth in experimental models [71].

Additionally, IKK $\alpha$  phosphorylates N-CoR, akin to SMRT, facilitating its nuclear export from CRC cells [72]. The active nuclear IKK $\alpha$  isoform, IKK $\alpha$ (p45), is crucial for preventing apoptosis and thereby fostering tumor growth, specifically in HCT116 cells. Mechanistically, the association between active TAK1, BRAF, a complex containing IKK $\alpha$ (p45), and NEMO leads to SMRT and Histone H3 phosphorylation, which is vital for BRAF-mediated transformation independent from NF- $\kappa$ B signaling [73].

In keratinocytes, evidence demonstrates IKK $\alpha$ 's involvement in cancer initiation independently of NF- $\kappa$ B [74]. The deletion of IKK $\alpha$  induces skin squamous cell carcinoma in mice, affecting 14-3-3 $\sigma$  expression and prompting aberrant cell proliferation, disrupting skin homeostasis, and promoting cell transformation [75]. Additional studies support IKK $\alpha$ 's tumor suppressor role in the skin, linking its activity to the transforming growth factor beta (TGF $\beta$ ) pathway. Moreover, a specific variant of nuclear IKK $\alpha$  in keratinocytes leads to more aggressive tumors upon exposure to chemical carcinogens [76,77].

Basal cell carcinomas (BCCs) are the most prevalent among human cancers affecting the skin [78]. While the noncanonical NF- $\kappa$ B pathway relies on IKK $\alpha$ , its specific role in BCC remains unclear. One study indicated that, within both BCC and non-malignant conditions, IKK $\alpha$  is present in the nucleus. Within BCC, the nuclear IKK $\alpha$  directly interacts with the promoters of inflammation factors and LGR5, a marker for stem cells. This interaction leads to an increase in LGR5 expression through the activation of the STAT3

signaling pathway, thereby contributing to cancer progression. The activation of the STAT3 pathway influences the LGR5 expression in a manner dependent on IKK $\alpha$ , as demonstrated by the interplay between STAT3 and IKK $\alpha$ . Moreover, suppressing the IKK $\alpha$  impedes the tumor growth and transition from the epithelial stage to the mesenchymal stage. This finding highlights IKK $\alpha$ 's role as a genuine chromatin regulator in BCC. Its heightened expression facilitates oncogenic transformation by promoting the expression of genes related to stemness and inflammation. Consequently, these findings offer a fresh perspective on how IKK $\alpha$  may participate in the progression of BCC tumors within an inflammatory microenvironment [79].

Moreover, in a study by Mahato and colleagues [80], they have shown that the suppression of IKK $\alpha$  in prostate cancer cells using synthetic siRNAs affects tumor cell growth and invasiveness. In this study, the authors designed three synthetic siRNAs targeting the specific regions of IKK $\alpha$  mRNA and evaluated their ability to silence IKK $\alpha$  in PC-3 and DU145 cells. A range of assays, including wound healing, migration, proliferation, and cell cycle analysis, were employed to investigate how IKK $\alpha$  siRNAs biologically impacted prostate cancer cells. Interestingly, their results uncovered potent siRNAs that could silence IKK $\alpha$  by up to 70%, resulting in decreased wound healing, migration, invasion, and cell attachment capabilities in prostate cancer cells. Additionally, this study observed comparable anti-invasive effects in the presence of RANKL. However, silencing IKK $\alpha$  had minimal effects on cell proliferation and cell cycle distribution. These findings strongly indicate that IKK $\alpha$  significantly influences prostate cancer invasion and metastasis while playing a minor role in cell proliferation. Targeting IKK $\alpha$  using siRNA emerges as a promising therapeutic approach for managing prostate cancer by reducing invasion and metastasis without directly impacting cell proliferation [81,82].

Moreover, IKK $\alpha$  contributes to progesterone-induced tumor promotion in breast cancer, downstream of RANKL induction, and fosters metastatic spread relying on RANKL produced by tumor-infiltrating regulatory T cells. It phosphorylates Estrogen Receptor  $\alpha$ , its coactivator AIB1/SRC3, and induces targets like cyclin D1 and c-myc, driving breast cancer cell proliferation [83]. Clinical observations link IKK $\alpha$  expression in breast cancer cells with patient outcomes regardless of cellular localization. In triple-negative breast cancer (TNBC) cells, IKK $\alpha$  mediates Notch signaling triggered by the Notch ligand Jagged1, a pivotal pathway for TNBC Cancer Stem Cell survival [84]. Combining therapies targeting the intersection of Notch, AKT, and NF- $\kappa$ B pathways holds promise for therapeutic applications against cancer stem cells in TNBC [85].

#### 4.2. IKK $\beta$ (Inhibitor of $\kappa$ B Kinase Beta)

In contrast to IKK $\alpha$ , IKK $\beta$  is predominantly associated with the canonical NF- $\kappa$ B pathway. One of its key functions is the phosphorylation of I $\kappa$ B $\alpha$  and I $\kappa$ B $\beta$ . This phosphorylation event marks I $\kappa$ B proteins for degradation, allowing the release of p50-RelA dimers [86]. The degradation of I $\kappa$ B proteins leads to the liberation of NF- $\kappa$ B, a transcription factor crucial in orchestrating the expression of pro-inflammatory genes. This includes genes responsible for cytokine and chemokine production, thus fostering a sustained and amplified inflammatory response within the tumor microenvironment [87]. This persistent inflammation, driven by IKK $\beta$ , creates a milieu that nurtures tumor growth, angiogenesis, and metastasis [88].

The released dimers subsequently move into the nucleus, where they commence the transcription of genes linked with the canonical NF- $\kappa$ B pathway. Notably, the activation of IKK $\beta$  is frequently prompted by proinflammatory stimuli, connecting it to persistent inflammation [89]. This linkage further underscores IKK $\beta$ 's significance in fostering tumor progression, the formation of new blood vessels (angiogenesis), and the spread of cancer to distant sites (metastasis), highlighting its critical role in the context of cancer. However, it is important to note that the outcome of this inflammatory response is context-dependent, either influencing the promotion of tumor formation or the initiation of an immune reaction against tumors [90].

To date, diverse chemical inhibitors targeting IKK $\beta$  have been discovered, each employing distinct mechanisms [91] (see Table 2). Most of these inhibitors mimic ATP, displaying reversible, ATP-competitive behavior, often exhibiting some preference for inhibiting IKK $\beta$  over IKK $\alpha$  and other kinases [92]. Yet, due to the structural similarity of protein kinase ATP-binding sites, these ATP mimics can inadvertently affect other kinases, causing unintended effects at concentrations required to inhibit their primary target in cells [93]. Specifically, some commonly used ‘specific’ IKK $\beta$  inhibitors, such as Bay 11-7082 and TPCA-1, have been found to induce significant off-target effects [94]. For instance, Bay 11-7082 disrupts NF- $\kappa$ B by irreversibly deactivating the E2-conjugating enzymes Ubc13 and UbcH7, as well as the E3-ligase LUBAC, rather than directly inhibiting IKK activity. Similarly, TPCA-1 hampers the STAT3 signaling by directly binding to the STAT3 Src Homology 2 (SH2) domain, alongside its IKK $\beta$  inhibitory activity [95].

**Table 2.** The selected compounds of natural and synthetic origin that were investigated for their effects on IKK $\beta$  in various cancer cell lines. They modulate IKK $\beta$  activity, leading to a suppression of NF- $\kappa$ B signaling, and the consequent downregulation of genes associated with inflammation, cell survival, and proliferation.

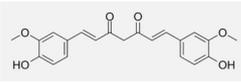
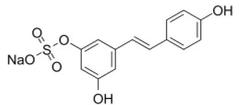
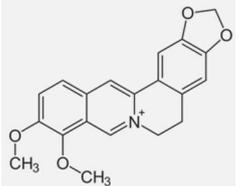
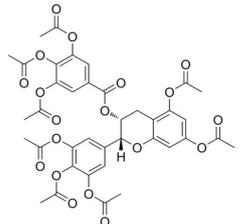
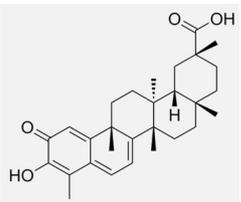
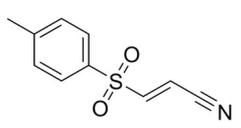
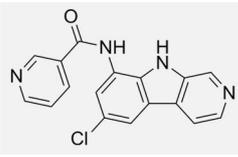
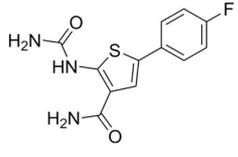
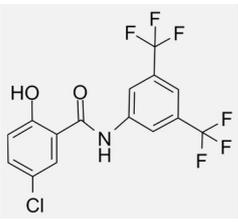
Compound Name	Source or Synthesis	Cell Line/Organism	Concentration ( $\mu$ M)	Incubation Time (h)	Observed Effect on IKK $\beta$ /Target	Structure
Curcumin [96]	Turmeric (Plant)	Various cancer cell lines (e.g., MCF-7, A549)	10–50	12–48	Inhibition of IKK $\beta$ phosphorylation and NF- $\kappa$ B activation, leading to reduced pro-inflammatory and pro-survival gene expression.	
Resveratrol [97]	Red grapes (plant)	Human prostate cancer cells (e.g., PC-3)	50–100	24–72	Suppression of IKK $\beta$ activity, resulting in reduced NF- $\kappa$ B-mediated transcription and anti-proliferative effects.	
Berberine [98]	Berberis pant	Various cancer cell lines (e.g., HCT-116, MDA-MB-231)	10–100	12–48	Inhibition of IKK $\beta$ phosphorylation, blocking NF- $\kappa$ B activation, and reducing the expression of pro-inflammatory and anti-apoptotic genes.	
EGCG (epigallocatechin-3-gallate) [99]	Green tea (plant)	Various cancer cell lines (e.g., A549, HCT-116)	20–100	24–48	Suppression of IKK $\beta$ phosphorylation, leading to decreased NF- $\kappa$ B activity and inhibition of pro-survival and pro-inflammatory pathways.	
Celastrol [100]	Thunder of god vine (plant)	Human breast cancer cells (e.g., MDA-MB-231)	0.5–1	6–24	Inhibition of IKK $\beta$ activity, blocking NF- $\kappa$ B signaling, and promoting apoptosis in cancer cells.	
BAY 11-7082 [101]	Synthetic compound	Multiple cancer cell lines (e.g., HeLa, U87)	5–20	2–24	Direct inhibition of IKK $\beta$ activity, leading to the suppression of NF- $\kappa$ B signaling and the downregulation of pro-survival and pro-inflammatory genes.	

Table 2. Cont.

Compound Name	Source or Synthesis	Cell Line/Organism	Concentration ( $\mu\text{M}$ )	Incubation Time (h)	Observed Effect on IKK $\beta$ /Target	Structure
PS1145 [102]	Synthetic compound	Various cancer cell lines (e.g., A549, MDA-MB-231)	1–10	4–24	Selective inhibition of IKK $\beta$ , resulting in the attenuation of NF- $\kappa$ B signaling and the reduction in pro-inflammatory and anti-apoptotic gene expression.	
TPCA-1 [103]	Synthetic compound	Human lung cancer cells (e.g., H1299)	1–5	6–24	Inhibition of IKK $\beta$ kinase activity, leading to the suppression of NF- $\kappa$ B-mediated transcription and anti-proliferative effects.	
IMD-0354 [104]	Synthetic compound	Prostate cancer cells (e.g., PC-3)	10–50	6–48	Inhibition of IKK $\beta$ activity, resulting in reduced NF- $\kappa$ B signaling and the downregulation of genes associated with cell survival and inflammation.	

Currently, the most potent ATP-competitive inhibitors for IKK $\beta$  include MLN-120B and BI605906, showcasing over 50-fold and over 300-fold selectivity for IKK $\beta$  over IKK $\alpha$ , respectively [105].

Recent research indicates potential toxicity and side effects correlated with IKK $\beta$  inhibition, such as the onset of inflammatory skin diseases and the heightened vulnerability of colonic epithelium to various stressors [106,107]. Severe liver malfunction has been observed in mice with IKK $\beta$  deficiencies, and intestinal and liver toxicity has surfaced in numerous clinical trials involving IKK $\beta$  inhibitors, potentially restricting their clinical applicability [108].

## 5. The Bright Side: IKKs as Tumor Suppressors

### 5.1. IKK $\alpha$ in Tumor Suppression

Despite its role in promoting cancer in some contexts, IKK $\alpha$  also has tumor-suppressive functions. It can induce cellular senescence in response to oncogenic stress, causing cells to enter a state of irreversible growth arrest [109]. Cells that undergo senescence not only cease dividing but also release substances referred to as the senescence-associated secretory phenotype (SASP). These substances attract immune cells, which in turn play a role in eliminating cells that could potentially develop into tumors [110].

A significant association exists between DNA damage-triggered senescence and the NF- $\kappa$ B-regulated SASP [111]. When exposed to genotoxic stress, the Ataxia telangiectasia mutated (ATM) kinase activates NF- $\kappa$ B by triggering the post-translational modifications (PTMs) of NEMO, which play a pivotal role in NF- $\kappa$ B activation. NEMO activation by ATM subsequently triggers the IKK complex, culminating in the nuclear translocation of NF- $\kappa$ B and the transcription of numerous genes related to SASP [112]. In melanoma, senescent cells produce a secretome characterized by the pro-invasive and pro-tumorigenic properties, relying on PARP-1 and NF- $\kappa$ B [113].

The expression of SASP components IL-6 and IL-8 necessitates I $\kappa$ B $\zeta$  during both DNA damage-induced senescence and oncogene-induced senescence (OIS), establishing I $\kappa$ B $\zeta$  as a crucial modulator of the proinflammatory SASP [114].

Multiple strands of evidence indicate that the continuous DNA damage response (DDR) is crucial for robust SASP production. The depletion of DDR elements like ATM, NBS1, or CHK2 inhibits the expression of IL-6, IL-8, and several GRO family members [115].

Consequently, it has been demonstrated, at least in specific experimental setups, that DDR activation—not just the presence of DNA damage itself—governs senescent states and SASP regulation [116].

Metformin, an anti-diabetic medication with diverse effects, also exerts activity on senescent cells [117]. One of its effects involves impairing the SASP of RAS-induced senescent cells without impeding proliferative arrest. This occurs through the inhibition of IKK $\alpha$ / $\beta$  and I $\kappa$ B phosphorylation by metformin, preventing the nuclear translocation of p65 (RelA) [118]. Metformin negatively influences NF- $\kappa$ B without affecting other inflammatory pathways like p38, JNK, and IRF. The inhibition of SASP by metformin might contribute to the observed anti-aging effects post-metformin treatment [119].

### 5.2. IKK $\beta$ in Antitumor Immune Responses

One of the central mechanisms through which IKK $\beta$  contributes to antitumor immunity is the activation of immune cells, particularly T cells and dendritic cells (DCs) [2]. These immune cells play pivotal roles in orchestrating immune responses against cancer.

IKK $\beta$  activation in T cells enhances their responsiveness and effector functions. T cells are the foot soldiers of the immune system, responsible for recognizing and eliminating cancer cells [120]. The activation of the IKK $\beta$ /NF- $\kappa$ B pathway in T cells augments their activation and proliferation. This, in turn, leads to an increased pool of cytotoxic T lymphocytes (CTLs) that can effectively target and kill cancer cells [121]. Additionally, activated T cells can infiltrate the tumor microenvironment, exerting their antitumor effects directly at the site of malignancy.

T cells capable of recognizing tumor-associated antigens exhibit potential in eradicating tumors [122]. Despite their presence in cancer patients—both in circulation and within tumors—these tumor-reactive T cells often fail to prevent tumor progression over time, indicating a probable decline in their functional abilities [123]. The direct analysis of these tumor antigen-specific T cells revealed deficiencies in cytokine production and cytolytic activity. Efforts to intervene and restore T cell function have shown promise in clinical settings but frequently result in partial responses. Understanding the mechanisms underlying T cell dysfunction in cancer remains crucial for enhancing therapeutic efficacy [124].

One critical pathway for T cell function involves the activation of I $\kappa$ B kinase  $\beta$  (IKK $\beta$ ) and the subsequent activation of NF- $\kappa$ B. In the tumor environment, T cell-NF- $\kappa$ B activity is often hindered, leading to reduced functionality in T cells isolated from cancer patients [125]. Recent studies conducted with mouse models exhibiting compromised NF- $\kappa$ B downstream of the T cell receptor (TCR) underscored the critical importance of T cell-NF- $\kappa$ B activation in the release of cytokines, specific targeting, and the destruction of antigens, and the *in vivo* eradication of tumors [126]. This indicates that reduced T cell-NF- $\kappa$ B activity induced by growing tumors compromises anti-tumor T cell responses, fostering a cycle favoring tumor growth. Consequently, exploring methods to stimulate T cell-intrinsic NF- $\kappa$ B activity becomes a compelling avenue for enhancing anti-tumor immunity [127,128].

In a study by Evaristo et al. [129], novel genetic mouse models expressing constitutively active IKK $\beta$  (caIKK $\beta$ ) specifically in T cells were employed. The results demonstrated that the T cell-specific expression of caIKK $\beta$  significantly improved tumor control, even in cases of established tumors. Thus, stimulating T cell-intrinsic NF- $\kappa$ B appears crucial in responding to cancer growth, suggesting that the therapeutic manipulation of the IKK $\beta$ /NF- $\kappa$ B axis holds promise for boosting anti-tumor immune responses.

DCs are antigen-presenting cells that play a critical role in initiating and shaping antitumor immune responses. IKK $\beta$  activation in DCs enhances their ability to capture, process, and present tumor antigens to T cells [130]. This process, known as antigen presentation, is a crucial step in initiating an adaptive immune response against cancer. The activation of the IKK $\beta$ /NF- $\kappa$ B pathway in DCs results in the upregulated expression of co-stimulatory molecules and cytokines that are necessary for efficient T-cell priming [131].

IKK $\beta$  activation in DCs leads to the upregulation of co-stimulatory molecules, such as CD80 and CD86 [132]. These molecules interact with their corresponding receptors on

T cells, providing essential co-stimulatory signals that are required for T-cell activation and proliferation. The enhanced expression of these co-stimulatory molecules by IKK $\beta$ -activated DCs amplifies the effectiveness of T-cell priming and the subsequent antitumor immune response [133].

Baratin et al., 2015 [134] conducted a comparative analysis of the transcriptomes of NLT-DCs within the skin and their migratory counterparts located in the draining lymph nodes (LNs). Through this investigation, they identified a novel gene network that is regulated by the NF- $\kappa$ B pathway and is specific to migratory dendritic cells. Their findings demonstrate that the targeted deletion of IKK $\beta$ , a key activator of NF- $\kappa$ B, in dendritic cells, hampers the accumulation of NLT-DCs in LNs and impairs the conversion of regulatory T cells in vivo. These outcomes are closely associated with disruptions in immune tolerance and the onset of autoimmune responses.

The activation of IKK $\beta$  leads to a heightened production of proinflammatory cytokines like interleukin-12 (IL-12) and interferon-gamma (IFN- $\gamma$ ) by DCs [135]. These cytokines serve a pivotal function in fostering an immune environment that supports anti-tumor immunity. IL-12, for instance, can skew the immune response towards a Th1 phenotype, characterized by enhanced cytotoxic activity and IFN- $\gamma$  production by T cells. IFN- $\gamma$ , on the other hand, has direct antitumor effects and can activate other immune cells to contribute to tumor eradication [136].

## 6. Distinct Phosphorylation Events and Kinases

Serine/threonine phosphorylation by IKK $\alpha$  and IKK $\beta$  plays a pivotal role in regulating the NF- $\kappa$ B pathway. When NF- $\kappa$ B is inactive, it is typically sequestered in the cytoplasm by inhibitor proteins known as I $\kappa$ B (Inhibitor of  $\kappa$ B) [137]. The phosphorylation of serine residues within I $\kappa$ B proteins, especially I $\kappa$ B $\alpha$  and I $\kappa$ B $\beta$ , is a critical event orchestrated by IKK $\beta$ . This phosphorylation serves as a recognition signal for the E3 ubiquitin ligase, which ubiquitinates I $\kappa$ B $\alpha$  and I $\kappa$ B $\beta$ e [138].

Tyrosine-phosphorylated STAT dimers represent the culmination of these intricate processes. Once formed, these dimers are ready to exert their transcriptional influence. STAT dimers translocate from the cytoplasm into the cell nucleus. Inside the nucleus, they bind to specific DNA sequences known as enhancer elements or response elements in the regulatory regions of target genes [139]. This binding event is specific to the dimer's composition and the cytokine signals received.

The binding of tyrosine-phosphorylated STAT dimers to these regulatory elements serves as a molecular switch, initiating the transcription of genes associated with immune responses and inflammatory processes [140]. The regulated genes often encode critical immune effectors, signaling molecules, and cytokines, shaping the cell's response and ultimately contributing to the immune and inflammatory outcomes observed in response to cytokine stimulation. Importantly, the activation of tyrosine kinases, such as JAKs, indirectly stimulates the NF- $\kappa$ B pathway [141]. This occurs through the cooperative efforts of transcription factors like STATs, which activate the gene expression related to immune responses. These genes may include those encoding proinflammatory cytokines and chemokines [142].

The coordination of various signaling pathways, involving both tyrosine phosphorylation and serine/threonine phosphorylation events, ensures that the cellular response is robust, efficient, and finely tuned to the needs of the immune system [143]. For instance, in viral infection, the release of interferons triggers the activation of JAK-STAT signaling, leading to the transcription of antiviral genes [144]. At the same time, the activation of the NF- $\kappa$ B pathway stimulates the production of proinflammatory cytokines, which, in turn, serve to attract immune cells to the site of infection. The interplay between these signaling pathways optimizes the host's response to the viral threat, underscoring the complexity of cross-talk mechanisms in immune regulation [145].

In the non-canonical NF- $\kappa$ B activation pathway, IKK $\alpha$  primarily phosphorylates a different substrate, p100. This phosphorylation leads to a unique processing event that is crucial for the activation of non-canonical NF- $\kappa$ B [146].

The phosphorylation of p100 by IKK $\alpha$  is followed by its partial proteasomal processing. This processing event results in the generation of a smaller protein fragment, p52. Importantly, p52 contains the DNA-binding domain necessary for transcriptional activity, allowing it to function as an NF- $\kappa$ B transcription factor [147]. The p52 subunit combines with other proteins, like RelB, to create dimers that then move into the nucleus. This sequence of events triggers the activation of the non-canonical NF- $\kappa$ B pathway, which functions differently compared to the canonical pathway [148].

The mechanism behind the phosphorylation of the I $\kappa$ B kinase T-loop remains a significant unanswered query. Suggestions have arisen proposing the involvement of IKK kinases (IKKKs) in this process, drawing an analogy to other signaling pathways [149]. One prominent example is TAK1, known for its involvement in the JNK pathway [150]. In cell-free assays, TAK1, along with the adaptor proteins TAB 1 and TAB 2, has been identified as a TRAF6-regulated IKK activator [151]. TAB 2's role in recruiting TAK1 to the K63-linked polyubiquitin chains of upstream regulators likely induces IKK $\beta$  phosphorylation through proximity-driven mechanisms [152]. However, TAK1 is not a universal IKKK but rather functions as a regulatory module impacting the IKK activation based on stimuli and cell type. Another potential IKKK, MEKK3 [153], has been proposed as it can phosphorylate IKK in vitro, and its deficiency correlates with the reduced NF- $\kappa$ B activation in response to various stimulations like TNF, IL-1, or TLR. IL-1-induced NF- $\kappa$ B activation has been linked to MEKK3 alongside TAK1 [154]. However, it is also suggested that IKK subunits might undergo activation through trans-autophosphorylation rather than via an IKKK. Recent structural and composition analyses support these possibilities, indicating that trans-autophosphorylation and IKKK-dependent phosphorylation might work sequentially or in parallel to achieve optimal kinase activation [155].

The dimeric NF- $\kappa$ B transcription factor, consisting of Rel family subunits that bind to DNA, plays a pivotal role in immune and inflammatory responses. NF- $\kappa$ B has recently been identified as a protector against apoptosis induced by tumor necrosis factor (TNF) and various genotoxic agents [156]. Typically, NF- $\kappa$ B dimers are confined within the cytoplasm due to their interaction with inhibitory I $\kappa$ B proteins. These proteins bind to the Rel homology domain (RHD), which is responsible for the dimerization, nuclear translocation, and DNA binding functions of NF- $\kappa$ B/Rel proteins [157]. When cells are stimulated by proinflammatory cytokines (e.g., IL-1, TNF), bacterial lipopolysaccharide (LPS) or phorbol ester (TPA), specific I $\kappa$ Bs (such as I $\kappa$ B $\alpha$  and I $\kappa$ B $\beta$ ) undergo rapid phosphorylation at certain N-terminal residues. This phosphorylation leads to their subsequent polyubiquitination and degradation via the 26S proteasome [158].

The breakdown of these inhibitory I $\kappa$ B proteins liberates the NF- $\kappa$ B dimer, enabling its migration into the nucleus to commence gene transcription [159]. Altering the phosphorylation sites on I $\kappa$ B $\alpha$  through specific mutations prevents its phosphorylation, ubiquitination, and subsequent degradation. Mutants like I $\kappa$ B $\alpha$ (A32/36) act as powerful inhibitors of NF- $\kappa$ B activation. Efforts to comprehend the regulation of this pathway have concentrated on identifying the responsible protein kinase(s). The I $\kappa$ B kinase (IKK) complex, a 900 kDa protein kinase complex, phosphorylates I $\kappa$ B $\alpha$  and I $\kappa$ B $\beta$  at sites crucial for their ubiquitination and degradation [160]. Another protein kinase complex phosphorylating I $\kappa$ B has been described, but its relationship with IKK remains unclear. IKK's activity is rapidly stimulated by IL-1 or TNF and is dependent on its phosphorylation [161]. One component of the IKK complex, IKK $\alpha$ , an 85 kDa polypeptide containing a protein kinase domain and protein interaction motifs, has been molecularly identified. IKK $\alpha$ 's expression is vital for NF- $\kappa$ B activation by various stimuli. IKK $\alpha$  has been isolated as a NIK-interacting protein, suggesting its role in NIK-mediated NF- $\kappa$ B activation [162]. While a catalytically inactive IKK $\alpha$  mutant can inhibit NF- $\kappa$ B activation, it can still interact with other IKK components to form a functional but less active I $\kappa$ B kinase complex.

Ubiquitination is a post-translational modification that tags proteins for degradation by the proteasome [163]. Following ubiquitination, I $\kappa$ B $\alpha$  and I $\kappa$ B $\beta$  are targeted for proteasomal degradation. As a result, they are rapidly degraded, freeing NF- $\kappa$ B from its sequestration. The released NF- $\kappa$ B transcription factors can subsequently migrate to the cell nucleus and initiate the transcription of genes [164].

The activation of IKKs appears to hinge on induced proximity from densely organized signaling complexes and the binding of adapter proteins like NEMO or TAB proteins. Non-degradative polyubiquitination, essential for IKK complex activation, triggers these processes [165].

TRAF6 stands as the initial ubiquitin E3 ligase identified, catalyzing K63-linked auto-ubiquitination alongside Ubc13 and Uev1A, subsequently initiating IKK activation. TRAF6 participates in numerous NF- $\kappa$ B-stimulating signaling pathways, including those triggered by IL-1R, TLR, TCR, RIG-I-like receptor, and DNA double-strand breaks [166]. In IL-1 signaling, TRAF6's enzymatic activity, rather than auto-ubiquitination, is essential for NF- $\kappa$ B activation. TRAF6 not only self-ubiquitinates but also mediates the K63-linked polyubiquitination of various pathway components like IRAK1, MALT1, and TAK1 [167]. Moreover, it is suggested that TRAF6 generates free, unanchored K63-linked polyubiquitin, serving as a docking platform in IKK activation [168]. Several K63-specific E3 ligases involved in distinct NF- $\kappa$ B signaling cascades were identified, such as TRAF2/5, or TRIM25 in the TNFR, IL-1R/TLR, and RIG-I pathways. Additionally, several proposed NF- $\kappa$ B pathway regulators are substrates of inducible K63 ubiquitination, including Bcl10, NOD2, and ELKS [169].

TNF $\alpha$  signaling does not rely on K63-linked ubiquitination, suggesting the importance of alternative non-degradative polyubiquitination in this pathway [170]. For instance, the linear, M1-linked ubiquitination of NEMO and RIP1 by the LUBAC complex is crucial for NF- $\kappa$ B activation. LUBAC is associated with the TNFR1 signaling complex. LUBAC-mediated M1-linked ubiquitination contributes to various NF- $\kappa$ B activations but is dispensable for B-cell receptor signaling. Mutations affecting LUBAC components were linked to immune-related disorders, showcasing their physiological relevance [171].

Multiple ubiquitin linkages seem to play roles in NF- $\kappa$ B signaling, adding to the complexity of ubiquitin-mediated processes [172]. E3 ligases like cIAP1 and TRIM23 catalyze distinct ubiquitin chain types, and certain proteins show modifications with various ubiquitin linkages in response to stimuli. Additionally, the mono-ubiquitination of proteins like NEMO has been shown to impact NF- $\kappa$ B activation. These diverse modifications coordinate specific protein interactions in NF- $\kappa$ B signaling pathways, although many intricacies regarding these actions remain undiscovered [173].

## 7. Genetic and Epigenetic Regulation

While primary genetic mutations within the I $\kappa$ B kinase genes themselves are relatively rare occurrences in the realm of cancer, modifications affecting their upstream regulatory elements can exert profound and far-reaching effects. Somatic mutations affecting lysine 171 within the IKBKB gene, responsible for encoding the critical activating kinase (IKK $\beta$ ) in the canonical NF $\kappa$ B signaling pathway, were observed in splenic marginal zone lymphomas and multiple myeloma. Lysine 171 is part of a positively charged pocket crucial for interaction with the activation loop phosphate in the naturally activated kinase [174].

Their findings demonstrate that both K171E IKK $\beta$  and K171T IKK $\beta$  variants function as kinases that remain constantly active, even in the absence of activation loop phosphorylation. Through predictive modeling and biochemical investigations, we elucidate why mutations in a positively charged residue within the cationic pocket of a kinase reliant on activation loop phosphorylation lead to persistent activation [175].

Utilizing transcription activator-like effector nuclease (TALEN)-based knock-in mutagenesis, we present evidence from a B lymphoid context indicating the involvement of K171E IKK $\beta$  in the development of lymphomas. Genetic mutations in TRAF proteins have the potential to engender the persistent and aberrant activation of the NF- $\kappa$ B pathway, a

phenomenon observed in various cancer types [176]. A notable instance is the frequent occurrence of mutations in the TRAF3 gene within the context of multiple myeloma, which leads to a chronic state of NF- $\kappa$ B pathway activation. The discernment of such genetic anomalies, often made possible through advanced techniques like whole-genome sequencing, not only advances our comprehension of the multifaceted terrain of cancer biology but also unveils promising therapeutic targets for potential intervention [177].

Multiple mechanisms contribute to NF- $\kappa$ B transcriptional regulation beyond its binding to  $\kappa$ B regulatory elements in DNA. In non-neuronal cells, the signaling elements of the NF- $\kappa$ B pathway participate in gene expression control through histone phosphorylation and acetylation in coordination with histone deacetylases (HDACs). The I $\kappa$ B protein variant, I $\kappa$ B $\alpha$ , independently governs transcription by engaging with HDAC1 and HDAC3. Additionally, the IKK $\alpha$  subunit operates distinctly from the IKK complex, influencing the cytokine-induced gene expression by modulating histone H3 phosphorylation [178]. These investigations reveal the novel functions of NF- $\kappa$ B signaling components, like I $\kappa$ B $\alpha$  and IKK $\alpha$ , in autonomously regulating the chromatin structure and gene expression from NF- $\kappa$ B's direct DNA binding.

Previously formed memories are prone to disruption immediately post-recall, necessitating reconsolidation. While protein translation mechanisms are acknowledged for their role in memory reconsolidation, research into gene transcription mechanisms remains relatively limited in this context [179].

An interesting study [180] indicated that the retrieval of contextual conditioned fear memories activates the NF- $\kappa$ B pathway, regulating histone H3 phosphorylation and acetylation at specific gene promoters in the hippocampus, particularly mediated by IKK $\alpha$  rather than the NF- $\kappa$ B DNA-binding complex. Behaviorally, inhibiting IKK $\alpha$ 's control over a chromatin structure or NF- $\kappa$ B DNA-binding complex activity results in impairments in fear memory reconsolidation. Elevated histone acetylation offsets this memory deficit when faced with the IKK blockade. These results offer new insights into IKK-mediated transcriptional mechanisms in the hippocampus essential for memory reconsolidation.

MicroRNAs (miRNAs) are another essential post-transcriptional regulator of I $\kappa$ B kinases [181]. One example is miR-21, a well-known oncogenic miRNA that plays a role in regulating the NF- $\kappa$ B pathway in various cancers. In certain scenarios, miR-21 targets PTEN (Phosphatase and Tensin Homolog), an inhibitor of the NF- $\kappa$ B pathway. This targeting leads to increased NF- $\kappa$ B activity in cancer cells, which, in turn, promotes cell survival, proliferation, and resistance to apoptosis [182].

## 8. Conclusions

I $\kappa$ B kinases, IKK $\alpha$  and IKK $\beta$ , represent a multifaceted and pivotal aspect of cancer biology, serving as central players in inflammation, immune responses, and tumorigenesis. Their dual role in cancer, which is shaped by myriad factors including distinct phosphorylation events, genetic and epigenetic regulation, and therapeutic implications, underscores the intricacies of their functions.

The potential of I $\kappa$ B kinases as therapeutic targets in cancer therapy is a burgeoning field. Precision medicine, which aims to individualize the treatment strategies based on genetic and molecular profiles, is at the forefront of this effort. While challenges such as resistance to IKK inhibitors persist, the development of combinatorial therapies and immunomodulation strategies offers hope for overcoming these obstacles.

**Funding:** This research received no external funding.

**Conflicts of Interest:** The authors declare no conflict of interest.

## References

1. Mitchell, S.; Vargas, J.; Hoffmann, A. Signaling via the NF $\kappa$ B system. *Wiley Interdiscip. Rev. Syst. Biol. Med.* **2016**, *8*, 227–241. [[CrossRef](#)] [[PubMed](#)]
2. Mulero, M.C.; Huxford, T.; Ghosh, G. NF- $\kappa$ B, I $\kappa$ B, and IKK: Integral Components of Immune System Signaling. *Adv. Exp. Med. Biol.* **2019**, *1172*, 207–226. [[CrossRef](#)] [[PubMed](#)]
3. Liu, F.; Xia, Y.; Parker, A.S.; Verma, I.M. IKK biology. *Immunol. Rev.* **2012**, *246*, 239–253. [[CrossRef](#)] [[PubMed](#)]
4. Durand, J.K.; Baldwin, A.S. Targeting IKK and NF- $\kappa$ B for Therapy. *Adv. Protein Chem. Struct. Biol.* **2017**, *107*, 77–115. [[CrossRef](#)] [[PubMed](#)]
5. Antonia, R.J.; Hagan, R.S.; Baldwin, A.S. Expanding the View of IKK: New Substrates and New Biology. *Trends Cell Biol.* **2021**, *31*, 166–178. [[CrossRef](#)]
6. Senegas, A.; Gautheron, J.; Maurin, A.G.; Courtois, G. IKK-related genetic diseases: Probing NF- $\kappa$ B functions in humans and other matters. *Cell. Mol. Life Sci.* **2015**, *72*, 1275–1287, Erratum in *Cell. Mol. Life Sci.* **2015**, *72*, 1289. [[CrossRef](#)] [[PubMed](#)]
7. Huang, W.C.; Hung, M.C. Beyond NF- $\kappa$ B activation: Nuclear functions of I $\kappa$ B kinase  $\alpha$ . *J. Biomed. Sci.* **2013**, *20*, 3. [[CrossRef](#)]
8. Gu, L.; Zhu, Y.; Lin, X.; Lu, B.; Zhou, X.; Zhou, F.; Zhao, Q.; Prochownik, E.V.; Li, Y. The IKK $\beta$ -USP30-ACLY Axis Controls Lipogenesis and Tumorigenesis. *Hepatology* **2021**, *73*, 160–174. [[CrossRef](#)]
9. Al Hamrashdi, M.; Brady, G. Regulation of IRF3 activation in human antiviral signaling pathways. *Biochem. Pharmacol.* **2022**, *200*, 115026. [[CrossRef](#)]
10. Fitzgerald, K.A.; McWhirter, S.M.; Faia, K.L.; Rowe, D.C.; Latz, E.; Golenbock, D.T.; Coyle, A.J.; Liao, S.M.; Maniatis, T. IKKepsilon and TBK1 are essential components of the IRF3 signaling pathway. *Nat. Immunol.* **2003**, *4*, 491–496. [[CrossRef](#)]
11. Willems, M.; Dubois, N.; Musumeci, L.; Bours, V.; Robe, P.A. I $\kappa$ B $\zeta$ : An emerging player in cancer. *Oncotarget* **2016**, *7*, 66310–66322. [[CrossRef](#)] [[PubMed](#)]
12. Yu, Z.; Gao, J.; Zhang, X.; Peng, Y.; Wei, W.; Xu, J.; Li, Z.; Wang, C.; Zhou, M.; Tian, X.; et al. Characterization of a small-molecule inhibitor targeting NEMO/IKK $\beta$  to suppress colorectal cancer growth. *Signal Transduct. Target. Ther.* **2022**, *7*, 71, Erratum in *Signal Transduct. Target. Ther.* **2023**, *8*, 330. [[CrossRef](#)] [[PubMed](#)]
13. Zhu, F.; Hu, Y. Integrity of IKK/NF- $\kappa$ B Shields Thymic Stroma That Suppresses Susceptibility to Autoimmunity, Fungal Infection, and Carcinogenesis. *Bioessays* **2018**, *40*, e1700131. [[CrossRef](#)] [[PubMed](#)]
14. Courtois, G.; Israël, A. IKK regulation and human genetics. *Curr. Top. Microbiol. Immunol.* **2011**, *349*, 73–95. [[CrossRef](#)] [[PubMed](#)]
15. May, M.J.; Ghosh, S. Signal transduction through NF- $\kappa$ B. *Immunol. Today* **1998**, *19*, 80–88. [[CrossRef](#)] [[PubMed](#)]
16. Hellweg, C.E. The Nuclear Factor  $\kappa$ B pathway: A link to the immune system in the radiation response. *Cancer Lett.* **2015**, *368*, 275–289. [[CrossRef](#)] [[PubMed](#)]
17. Hayden, M.S.; Ghosh, S. Shared principles in NF-kappaB signaling. *Cell* **2008**, *132*, 344–362. [[CrossRef](#)]
18. Lawrence, T. The nuclear factor NF-kappaB pathway in inflammation. *Cold Spring Harb. Perspect. Biol.* **2009**, *1*, a001651. [[CrossRef](#)]
19. Poma, P. NF- $\kappa$ B and Disease. *Int. J. Mol. Sci.* **2020**, *21*, 9181. [[CrossRef](#)]
20. DiDonato, J.A.; Mercurio, F.; Karin, M. NF- $\kappa$ B and the link between inflammation and cancer. *Immunol. Rev.* **2012**, *246*, 379–400. [[CrossRef](#)]
21. Oeckinghaus, A.; Hayden, M.S.; Ghosh, S. Crosstalk in NF- $\kappa$ B signaling pathways. *Nat. Immunol.* **2011**, *12*, 695–708. [[CrossRef](#)] [[PubMed](#)]
22. Paul, A.; Edwards, J.; Pepper, C.; Mackay, S. Inhibitory- $\kappa$ B Kinase (IKK)  $\alpha$  and Nuclear Factor- $\kappa$ B (NF $\kappa$ B)-Inducing Kinase (NIK) as Anti-Cancer Drug Targets. *Cells.* **2018**, *7*, 176. [[CrossRef](#)] [[PubMed](#)]
23. Li, Q.; Verma, I.M. NF-kappaB regulation in the immune system. *Nat. Rev. Immunol.* **2002**, *2*, 725–734, Erratum in *Nat. Rev. Immunol.* **2002**, *2*, 975. [[CrossRef](#)] [[PubMed](#)]
24. Zhang, Q.; Lenardo, M.J.; Baltimore, D. 30 Years of NF- $\kappa$ B: A Blossoming of Relevance to Human Pathobiology. *Cell* **2017**, *168*, 37–57. [[CrossRef](#)] [[PubMed](#)]
25. Mirzaei, S.; Saghari, S.; Bassiri, F.; Raesi, R.; Zarrabi, A.; Hushmandi, K.; Sethi, G.; Tergaonkar, V. NF- $\kappa$ B as a regulator of cancer metastasis and therapy response: A focus on epithelial-mesenchymal transition. *J. Cell. Physiol.* **2022**, *237*, 2770–2795. [[CrossRef](#)] [[PubMed](#)]
26. Hayden, M.S.; West, A.P.; Ghosh, S. SnapShot: NF-kappaB signaling pathways. *Cell* **2006**, *127*, 1286–1287. [[CrossRef](#)] [[PubMed](#)]
27. Wibisana, J.N.; Okada, M. Encoding and decoding NF- $\kappa$ B nuclear dynamics. *Curr. Opin. Cell Biol.* **2022**, *77*, 102103. [[CrossRef](#)]
28. Gilmore, T.D.; Wolenski, F.S. NF- $\kappa$ B: Where did it come from and why? *Immunol. Rev.* **2012**, *246*, 14–35. [[CrossRef](#)]
29. Williams, L.M.; Gilmore, T.D. Looking Down on NF- $\kappa$ B. *Mol. Cell Biol.* **2020**, *40*, e00104-20. [[CrossRef](#)]
30. Pujari, R.; Hunte, R.; Khan, W.N.; Shembade, N. A20-mediated negative regulation of canonical NF- $\kappa$ B signaling pathway. *Immunol. Res.* **2013**, *57*, 166–171. [[CrossRef](#)]
31. Pham, A.M.; TenOever, B.R. The IKK Kinases: Operators of Antiviral Signaling. *Viruses* **2010**, *2*, 55–72. [[CrossRef](#)] [[PubMed](#)]
32. Zinatizadeh, M.R.; Schock, B.; Chalbatani, G.M.; Zarandi, P.K.; Jalali, S.A.; Miri, S.R. The Nuclear Factor Kappa B (NF- $\kappa$ B) signaling in cancer development and immune diseases. *Genes Dis.* **2020**, *8*, 287–297. [[CrossRef](#)]
33. Sun, S.C. The non-canonical NF- $\kappa$ B pathway in immunity and inflammation. *Nat. Rev. Immunol.* **2017**, *17*, 545–558. [[CrossRef](#)] [[PubMed](#)]
34. Sun, S.C. Non-canonical NF- $\kappa$ B signaling pathway. *Cell Res.* **2011**, *21*, 71–85. [[CrossRef](#)]
35. Yu, H.; Lin, L.; Zhang, Z.; Zhang, H.; Hu, H. Targeting NF- $\kappa$ B pathway for the therapy of diseases: Mechanism and clinical study. *Signal Transduct. Target. Ther.* **2020**, *5*, 209. [[CrossRef](#)] [[PubMed](#)]

36. Hou, Y.; Liang, H.; Rao, E.; Zheng, W.; Huang, X.; Deng, L.; Zhang, Y.; Yu, X.; Xu, M.; Mauceri, H.; et al. Non-canonical NF- $\kappa$ B Antagonizes STING Sensor-Mediated DNA Sensing in Radiotherapy. *Immunity* **2018**, *49*, 490–503.e4. [[CrossRef](#)] [[PubMed](#)]
37. Gilmore, T.D. Introduction to NF-kappaB: Players, pathways, perspectives. *Oncogene* **2006**, *25*, 6680–6684. [[CrossRef](#)]
38. Struzik, J.; Szulc-Dąbrowska, L. Manipulation of Non-canonical NF- $\kappa$ B Signaling by Non-oncogenic Viruses. *Arch. Immunol. Ther. Exp.* **2019**, *67*, 41–48. [[CrossRef](#)]
39. Meyerovich, K.; Ortis, F.; Cardozo, A.K. The non-canonical NF- $\kappa$ B pathway and its contribution to  $\beta$ -cell failure in diabetes. *J. Mol. Endocrinol.* **2018**, *61*, F1–F6. [[CrossRef](#)]
40. Lu, X.; Chen, Q.; Liu, H.; Zhang, X. Interplay Between Non-Canonical NF- $\kappa$ B Signaling and Hepatitis B Virus Infection. *Front. Immunol.* **2021**, *29*, 730684. [[CrossRef](#)]
41. Noort, A.R.; Tak, P.P.; Tas, S.W. Non-canonical NF- $\kappa$ B signaling in rheumatoid arthritis: Dr Jekyll and Mr Hyde? *Arthritis Res. Ther.* **2015**, *17*, 15. [[CrossRef](#)] [[PubMed](#)]
42. Morgan, D.; Garg, M.; Tergaonkar, V.; Tan, S.Y.; Sethi, G. Pharmacological significance of the non-canonical NF- $\kappa$ B pathway in tumorigenesis. *Biochim. Biophys. Acta Rev. Cancer* **2020**, *1874*, 188449. [[CrossRef](#)] [[PubMed](#)]
43. Trares, K.; Ackermann, J.; Koch, I. The canonical and non-canonical NF- $\kappa$ B pathways and their crosstalk: A comparative study based on Petri nets. *Biosystems* **2022**, *211*, 104564. [[CrossRef](#)] [[PubMed](#)]
44. Tao, L.; Ren, X.; Zhai, W.; Chen, Z. Progress and Prospects of Non-Canonical NF- $\kappa$ B Signaling Pathway in the Regulation of Liver Diseases. *Molecules* **2022**, *27*, 4275. [[CrossRef](#)] [[PubMed](#)]
45. O'Donnell, A.; Pepper, C.; Mitchell, S.; Pepper, A. NF- $\kappa$ B and the CLL microenvironment. *Front. Oncol.* **2023**, *30*, 1169397. [[CrossRef](#)] [[PubMed](#)]
46. Shen, R.R.; Hahn, W.C. Emerging roles for the non-canonical IKKs in cancer. *Oncogene* **2011**, *30*, 631–641. [[CrossRef](#)] [[PubMed](#)]
47. Pflug, K.M.; Sitcheran, R. Targeting NF- $\kappa$ B-Inducing Kinase (NIK) in Immunity, Inflammation, and Cancer. *Int. J. Mol. Sci.* **2020**, *21*, 8470. [[CrossRef](#)]
48. Gu, M.; Zhou, X.; Sohn, J.H.; Zhu, L.; Jie, Z.; Yang, J.Y.; Zheng, X.; Xie, X.; Yang, J.; Shi, Y.; et al. NF- $\kappa$ B-inducing kinase maintains T cell metabolic fitness in antitumor immunity. *Nat. Immunol.* **2021**, *22*, 193–204, Erratum in Nat Immunol. 2021 Feb 11. [[CrossRef](#)]
49. Xiao, P.; Takiishi, T.; Violato, N.M.; Licata, G.; Dotta, F.; Sebastiani, G.; Marselli, L.; Singh, S.P.; Sze, M.; Van Loo, G.; et al. NF- $\kappa$ B-inducing kinase (NIK) is activated in pancreatic  $\beta$ -cells but does not contribute to the development of diabetes. *Cell Death Dis.* **2022**, *13*, 476. [[CrossRef](#)]
50. Cheng, J.; Feng, X.; Li, Z.; Zhou, F.; Yang, J.M.; Zhao, Y. Pharmacological inhibition of NF- $\kappa$ B-inducing kinase (NIK) with small molecules for the treatment of human diseases. *RSC Med. Chem.* **2021**, *12*, 552–565. [[CrossRef](#)]
51. Wang, B.; Shen, J. NF- $\kappa$ B Inducing Kinase Regulates Intestinal Immunity and Homeostasis. *Front. Immunol.* **2022**, *27*, 895636. [[CrossRef](#)] [[PubMed](#)]
52. Zhang, Z.; Zhong, X.; Shen, H.; Sheng, L.; Liangpunsakul, S.; Lok, A.S.; Omary, M.B.; Wang, S.; Rui, L. Biliary NIK promotes ductular reaction and liver injury and fibrosis in mice. *Nat. Commun.* **2022**, *13*, 5111. [[CrossRef](#)] [[PubMed](#)]
53. Choudhary, S.; Sinha, S.; Zhao, Y.; Banerjee, S.; Sathyanarayana, P.; Shahani, S.; Sherman, V.; Tilton, R.G.; Bajaj, M. NF-kappaB-inducing kinase (NIK) mediates skeletal muscle insulin resistance: Blockade by adiponectin. *Endocrinology* **2011**, *152*, 3622–3627. [[CrossRef](#)] [[PubMed](#)]
54. Thu, Y.M.; Su, Y.; Yang, J.; Splittgerber, R.; Na, S.; Boyd, A.; Mosse, C.; Simons, C.; Richmond, A. NF- $\kappa$ B inducing kinase (NIK) modulates melanoma tumorigenesis by regulating expression of pro-survival factors through the  $\beta$ -catenin pathway. *Oncogene* **2012**, *31*, 2580–2592. [[CrossRef](#)] [[PubMed](#)]
55. Fry, C.S.; Nayeem, S.Z.; Dillon, E.L.; Sarkar, P.S.; Tumurbaatar, B.; Urban, R.J.; Wright, T.J.; Sheffield-Moore, M.; Tilton, R.G.; Choudhary, S. Glucocorticoids increase skeletal muscle NF- $\kappa$ B inducing kinase (NIK): Links to muscle atrophy. *Physiol. Rep.* **2016**, *4*, e13014. [[CrossRef](#)]
56. Hahn, M.; Macht, A.; Waisman, A.; Hövelmeyer, N. NF- $\kappa$ B-inducing kinase is essential for B-cell maintenance in mice. *Eur. J. Immunol.* **2016**, *46*, 732–741. [[CrossRef](#)]
57. Chung, S.; Sundar, I.K.; Hwang, J.W.; Yull, F.E.; Blackwell, T.S.; Kinnula, V.L.; Bulger, M.; Yao, H.; Rahman, I. NF- $\kappa$ B inducing kinase, NIK mediates cigarette smoke/TNF $\alpha$ -induced histone acetylation and inflammation through differential activation of IKKs. *PLoS ONE* **2011**, *6*, e23488. [[CrossRef](#)]
58. Yamamoto, M.; Ito, T.; Shimizu, T.; Ishida, T.; Semba, K.; Watanabe, S.; Yamaguchi, N.; Inoue, J. Epigenetic alteration of the NF- $\kappa$ B-inducing kinase (NIK) gene is involved in enhanced NIK expression in basal-like breast cancer. *Cancer Sci.* **2010**, *101*, 2391–2397. [[CrossRef](#)]
59. Davis, J.L.; Thaler, R.; Cox, L.; Ricci, B.; Zannit, H.M.; Wan, F.; Faccio, R.; Dudakovic, A.; van Wijnen, A.J.; Veis, D.J. Constitutive activation of NF- $\kappa$ B inducing kinase (NIK) in the mesenchymal lineage using Osterix (Sp7)- or Fibroblast-specific protein 1 (S100a4)-Cre drives spontaneous soft tissue sarcoma. *PLoS ONE* **2021**, *16*, e0254426. [[CrossRef](#)]
60. Xiong, Y.; Torsoni, A.S.; Wu, F.; Shen, H.; Liu, Y.; Zhong, X.; Canet, M.J.; Shah, Y.M.; Omary, M.B.; Liu, Y.; et al. Hepatic NF- $\kappa$ B-inducing kinase (NIK) suppresses mouse liver regeneration in acute and chronic liver diseases. *Elife* **2018**, *2*, e34152. [[CrossRef](#)]
61. Shinzawa, M.; Konno, H.; Qin, J.; Akiyama, N.; Miyauchi, M.; Ohashi, H.; Miyamoto-Sato, E.; Yanagawa, H.; Akiyama, T.; Inoue, J. Catalytic subunits of the phosphatase calcineurin interact with NF- $\kappa$ B-inducing kinase (NIK) and attenuate NIK-dependent gene expression. *Sci. Rep.* **2015**, *1*, 10758. [[CrossRef](#)] [[PubMed](#)]

62. Vazquez-Santillan, K.; Melendez-Zajgla, J.; Jimenez-Hernandez, L.E.; Gaytan-Cervantes, J.; Muñoz-Galindo, L.; Piña-Sanchez, P.; Martinez-Ruiz, G.; Torres, J.; Garcia-Lopez, P.; Gonzalez-Torres, C.; et al. NF-kappaB-inducing kinase regulates stem cell phenotype in breast cancer. *Sci. Rep.* **2016**, *23*, 37340; Erratum in *Sci. Rep.* **2017**, *23*, 44971. [[CrossRef](#)] [[PubMed](#)]
63. Jiang, B.; Shen, H.; Chen, Z.; Yin, L.; Zan, L.; Rui, L. Carboxyl terminus of HSC70-interacting protein (CHIP) down-regulates NF- $\kappa$ B-inducing kinase (NIK) and suppresses NIK-induced liver injury. *J. Biol. Chem.* **2015**, *290*, 11704–11714. [[CrossRef](#)] [[PubMed](#)]
64. Pflug, K.M.; Lee, D.W.; Keeney, J.N.; Sitcheran, R. NF- $\kappa$ B-inducing kinase maintains mitochondrial efficiency and systemic metabolic homeostasis. *Biochim. Biophys. Acta Mol. Basis Dis.* **2023**, *1869*, 166682. [[CrossRef](#)] [[PubMed](#)]
65. Shen, H.; Ji, Y.; Xiong, Y.; Kim, H.; Zhong, X.; Jin, M.G.; Shah, Y.M.; Omary, M.B.; Liu, Y.; Qi, L.; et al. Medullary thymic epithelial NF- $\kappa$ B-inducing kinase (NIK)/IKK $\alpha$  pathway shapes autoimmunity and liver and lung homeostasis in mice. *Proc. Natl. Acad. Sci. USA* **2019**, *116*, 19090–19097. [[CrossRef](#)] [[PubMed](#)]
66. Thu, Y.M.; Richmond, A. NF- $\kappa$ B inducing kinase: A key regulator in the immune system and in cancer. *Cytokine Growth Factor Rev.* **2010**, *21*, 213–226. [[CrossRef](#)] [[PubMed](#)]
67. Shen, H.; Sheng, L.; Chen, Z.; Jiang, L.; Su, H.; Yin, L.; Omary, M.B.; Rui, L. Mouse hepatocyte overexpression of NF- $\kappa$ B-inducing kinase (NIK) triggers fatal macrophage-dependent liver injury and fibrosis. *Hepatology* **2014**, *60*, 2065–2076. [[CrossRef](#)]
68. Sheng, L.; Zhou, Y.; Chen, Z.; Ren, D.; Cho, K.W.; Jiang, L.; Shen, H.; Sasaki, Y.; Rui, L. NF- $\kappa$ B-inducing kinase (NIK) promotes hyperglycemia and glucose intolerance in obesity by augmenting glucagon action. *Nat. Med.* **2012**, *18*, 943–949. [[CrossRef](#)]
69. Hoberg, J.E.; Yeung, F.; Mayo, M.W. SMRT derepression by the IkappaB kinase alpha: A prerequisite to NF-kappaB transcription and survival. *Mol. Cell* **2004**, *16*, 245–255. [[CrossRef](#)]
70. Hoberg, J.E.; Popko, A.E.; Ramsey, C.S.; Mayo, M.W. IkappaB kinase alpha-mediated derepression of SMRT potentiates acetylation of RelA/p65 by p300. *Mol. Cell Biol.* **2006**, *26*, 457–471. [[CrossRef](#)]
71. Fernández-Majada, V.; Aguilera, C.; Villanueva, A.; Vilardell, F.; Robert-Moreno, A.; Aytés, A.; Real, F.X.; Capella, G.; Mayo, M.W.; Espinosa, L.; et al. Nuclear IKK activity leads to dysregulated notch-dependent gene expression in colorectal cancer. *Proc. Natl. Acad. Sci. USA* **2007**, *104*, 276–281, Erratum in *Proc. Natl. Acad. Sci. USA* **2008**, *105*, 5650. [[CrossRef](#)] [[PubMed](#)]
72. Fernández-Majada, V.; Pujadas, J.; Vilardell, F.; Capella, G.; Mayo, M.W.; Bigas, A.; Espinosa, L. Aberrant cytoplasmic localization of N-CoR in colorectal tumors. *Cell Cycle* **2007**, *6*, 1748–1752. [[CrossRef](#)] [[PubMed](#)]
73. Margalef, P.; Fernández-Majada, V.; Villanueva, A.; Garcia-Carbonell, R.; Iglesias, M.; López, L.; Martínez-Iniesta, M.; Villà-Freixa, J.; Mulero, M.C.; Andreu, M.; et al. A truncated form of IKK $\alpha$  is responsible for specific nuclear IKK activity in colorectal cancer. *Cell Rep.* **2012**, *2*, 840–854. [[CrossRef](#)] [[PubMed](#)]
74. Park, E.; Liu, B.; Xia, X.; Zhu, F.; Jami, W.B.; Hu, Y. Role of IKK $\alpha$  in skin squamous cell carcinomas. *Future Oncol.* **2011**, *7*, 123–134. [[CrossRef](#)] [[PubMed](#)]
75. Xie, Y.; Xie, K.; Gou, Q.; Chen, N. IkB kinase  $\alpha$  functions as a tumor suppressor in epithelial-derived tumors through an NF- $\kappa$ B-independent pathway (Review). *Oncol. Rep.* **2015**, *34*, 2225–2232. [[CrossRef](#)] [[PubMed](#)]
76. Liu, S.; Chen, Z.; Zhu, F.; Hu, Y. IkB kinase alpha and cancer. *J. Interferon Cytokine Res.* **2012**, *32*, 152–158. [[CrossRef](#)]
77. Zhu, F.; Park, E.; Liu, B.; Xia, X.; Fischer, S.M.; Hu, Y. Critical role of IkappaB kinase alpha in embryonic skin development and skin carcinogenesis. *Histol. Histopathol.* **2009**, *24*, 265–271. [[CrossRef](#)]
78. Tanese, K. Diagnosis and Management of Basal Cell Carcinoma. *Curr. Treat. Options Oncol.* **2019**, *20*, 13. [[CrossRef](#)]
79. Molin, S.C.; Grgic, M.; Ruzicka, T.; Herzinger, T. Silencing of the cell cycle checkpoint gene 14-3-3 $\sigma$  in basal cell carcinomas correlates with reduced expression of IKK- $\alpha$ . *J. Eur. Acad. Dermatol. Venereol.* **2014**, *28*, 1113–1116. [[CrossRef](#)]
80. Mahato, R.; Qin, B.; Cheng, K. Blocking IKK $\alpha$  expression inhibits prostate cancer invasiveness. *Pharm. Res.* **2011**, *28*, 1357–1369. [[CrossRef](#)]
81. Montes, M.; MacKenzie, L.; McAllister, M.J.; Roseweir, A.; McCall, P.; Hatzieremia, S.; Underwood, M.A.; Boyd, M.; Paul, A.; Plevin, R.; et al. Determining the prognostic significance of IKK $\alpha$  in prostate cancer. *Prostate* **2020**, *80*, 1188–1202. [[CrossRef](#)] [[PubMed](#)]
82. Nguyen, D.P.; Li, J.; Yadav, S.S.; Tewari, A.K. Recent insights into NF- $\kappa$ B signalling pathways and the link between inflammation and prostate cancer. *BJU Int.* **2014**, *114*, 168–176. [[CrossRef](#)] [[PubMed](#)]
83. Man, J.; Zhang, X. CUEDC2: An emerging key player in inflammation and tumorigenesis. *Protein Cell* **2011**, *2*, 699–703. [[CrossRef](#)] [[PubMed](#)]
84. Sau, A.; Lau, R.; Cabrita, M.A.; Nolan, E.; Crooks, P.A.; Visvader, J.E.; Pratt, M.A. Persistent Activation of NF- $\kappa$ B in BRCA1-Deficient Mammary Progenitors Drives Aberrant Proliferation and Accumulation of DNA Damage. *Cell Stem Cell* **2016**, *19*, 52–65. [[CrossRef](#)] [[PubMed](#)]
85. Liao, J.; Qin, Q.H.; Lv, F.Y.; Huang, Z.; Lian, B.; Wei, C.Y.; Mo, Q.G.; Tan, Q.X. IKK $\alpha$  inhibition re-sensitizes acquired adriamycin-resistant triple negative breast cancer cells to chemotherapy-induced apoptosis. *Sci. Rep.* **2023**, *13*, 6211. [[CrossRef](#)] [[PubMed](#)]
86. Ruocco, M.G.; Karin, M. IKK{beta} as a target for treatment of inflammation induced bone loss. *Ann. Rheum. Dis.* **2005**, *64* (Suppl. S4), iv81–iv85. [[CrossRef](#)]
87. Page, A.; Navarro, M.; Suárez-Cabrera, C.; Bravo, A.; Ramirez, A. Context-Dependent Role of IKK $\beta$  in Cancer. *Genes* **2017**, *8*, 376. [[CrossRef](#)]
88. Gan, J.; Guo, L.; Zhang, X.; Yu, Q.; Yang, Q.; Zhang, Y.; Zeng, W.; Jiang, X.; Guo, M. Anti-inflammatory therapy of atherosclerosis: Focusing on IKK $\beta$ . *J. Inflamm.* **2023**, *20*, 8. [[CrossRef](#)]

89. Zhang, G.; Li, J.; Purkayastha, S.; Tang, Y.; Zhang, H.; Yin, Y.; Li, B.; Liu, G.; Cai, D. Hypothalamic programming of systemic ageing involving IKK- $\beta$ , NF- $\kappa$ B and GnRH. *Nature* **2013**, *497*, 211–216. [[CrossRef](#)]
90. Zhou, P.; Hua, F.; Wang, X.; Huang, J.L. Therapeutic potential of IKK- $\beta$  inhibitors from natural phenolics for inflammation in cardiovascular diseases. *Inflammopharmacology* **2020**, *28*, 19–37. [[CrossRef](#)]
91. Heyninck, K.; Lahtela-Kakkonen, M.; Van der Veken, P.; Haegeman, G.; Vanden Berghe, W. Withaferin a inhibits NK-kappaB activation by targeting cysteine 179 in IKKbeta. *Biochem. Pharmacol.* **2014**, *91*, 501–509. [[CrossRef](#)]
92. Vanden Berghe, W.; Sabbe, L.; Kaileh, M.; Haegeman, G.; Heyninck, K. Molecular insight in the multifunctional activities of withaferin a. *Biochem. Pharmacol.* **2012**, *84*, 1282–1291. [[CrossRef](#)] [[PubMed](#)]
93. Kim, B.H.; Roh, E.; Lee, H.Y.; Lee, I.J.; Ahn, B.; Jung, S.H.; Lee, H.; Han, S.B.; Kim, Y. Benzoxathiole derivative blocks lipopolysaccharide-induced nuclear factor-kappaB activation and nuclear fac-tor-kappaB-regulated gene transcription through inactivating inhibitory kappaB kinase beta. *Mol. Pharmacol.* **2008**, *73*, 1309–1318. [[CrossRef](#)] [[PubMed](#)]
94. Dong, T.; Li, C.; Wang, X.; Dian, L.; Zhang, X.; Li, L.; Chen, S.; Cao, R.; Li, L.; Huang, N.; et al. Ainsliadimer a selectively inhibits IKK $\alpha$ / $\beta$  by covalently binding a conserved cysteine. *Nat. Commun.* **2015**, *6*, 6522. [[CrossRef](#)] [[PubMed](#)]
95. Prescott, J.A.; Cook, S.J. Targeting IKK $\beta$  in Cancer: Challenges and Opportunities for the Therapeutic Utilisation of IKK $\beta$  Inhibitors. *Cells* **2018**, *7*, 115. [[CrossRef](#)] [[PubMed](#)]
96. Chen, G.; Liu, S.; Pan, R.; Li, G.; Tang, H.; Jiang, M.; Xing, Y.; Jin, F.; Lin, L.; Dong, J. Curcumin Attenuates gp120-Induced Microglial Inflammation by Inhibiting Autophagy via the PI3K Pathway. *Cell. Mol. Neurobiol.* **2018**, *38*, 1465–1477. [[CrossRef](#)] [[PubMed](#)]
97. Yang, H.; Wang, Y.; Jin, S.; Pang, Q.; Shan, A.; Feng, X. Dietary resveratrol alleviated lipopolysaccharide-induced ileitis through Nrf2 and NF- $\kappa$ B signalling pathways in ducks (*Anas platyrhynchos*). *J. Anim. Physiol. Anim. Nutr.* **2022**, *106*, 1306–1320. [[CrossRef](#)]
98. Li, A.; Lin, C.; Xie, F.; Jin, M.; Lin, F. Berberine Ameliorates Insulin Resistance by Inhibiting IKK/NF- $\kappa$ B, JNK, and IRS-1/AKT Signaling Pathway in Liver of Gestational Diabetes Mellitus Rats. *Metab. Syndr. Relat. Disord.* **2022**, *20*, 480–488. [[CrossRef](#)]
99. Wheeler, D.S.; Catravas, J.D.; Odoms, K.; Denenberg, A.; Malhotra, V.; Wong, H.R. Epigallocatechin-3-gallate, a green tea-derived polyphenol, inhibits IL-1 beta-dependent proinflammatory signal transduction in cultured respiratory epithelial cells. *J. Nutr.* **2004**, *134*, 1039–1044. [[CrossRef](#)]
100. Veerappan, K.; Natarajan, S.; Ethiraj, P.; Vetrivel, U.; Samuel, S. Inhibition of IKK $\beta$  by celastrol and its analogues—An in silico and in vitro approach. *Pharm. Biol.* **2017**, *55*, 368–373. [[CrossRef](#)]
101. Rauert-Wunderlich, H.; Siegmund, D.; Maier, E.; Giner, T.; Bargou, R.C.; Wajant, H.; Stühmer, T. The IKK inhibitor Bay 11-7082 induces cell death independent from inhibition of activation of NF $\kappa$ B transcription factors. *PLoS ONE* **2013**, *8*, e59292. [[CrossRef](#)] [[PubMed](#)]
102. Lung, H.L.; Kan, R.; Chau, W.Y.; Man, O.Y.; Mak, N.K.; Fong, C.H.; Shuen, W.H.; Tsao, S.W.; Lung, M.L. The anti-tumor function of the IKK inhibitor PS1145 and high levels of p65 and KLF4 are associated with the drug resistance in nasopharyngeal carcinoma cells. *Sci. Rep.* **2019**, *9*, 12064. [[CrossRef](#)] [[PubMed](#)]
103. Sachse, F.; Becker, K.; Basel, T.J.; Weiss, D.; Rudack, C. IKK-2 inhibitor TPCA-1 represses nasal epithelial inflammation in vitro. *Rhinology* **2011**, *49*, 168–173. [[CrossRef](#)] [[PubMed](#)]
104. Waga, K.; Yamaguchi, M.; Miura, S.; Nishida, T.; Itai, A.; Nakanishi, R.; Kashiwakura, I. IKK $\beta$  Inhibitor IMD-0354 Attenuates Radiation Damage in Whole-body X-Irradiated Mice. *Oxid. Med. Cell. Longev.* **2019**, *27*, 5340290. [[CrossRef](#)] [[PubMed](#)]
105. Clark, K.; Peggie, M.; Plater, L.; Sorcek, R.J.; Young, E.R.; Madwed, J.B.; Hough, J.; McIver, E.G.; Cohen, P. Novel cross-talk within the IKK family controls innate immunity. *Biochem. J.* **2011**, *434*, 93–104. [[CrossRef](#)] [[PubMed](#)]
106. Li, Q.T.; Van Antwerp, D.; Mercurio, F.; Lee, K.F.; Verma, I.M. Severe liver degeneration in mice lacking the IkappaB kinase 2 gene. *Science* **1999**, *284*, 321–325. [[CrossRef](#)] [[PubMed](#)]
107. Lloná-Minguez, S.; Baiget, J.; Mackay, S.P. Small-molecule inhibitors of I $\kappa$ B kinase (IKK) and IKK-related kinases. *Pharm. Pat. Anal.* **2013**, *2*, 481–498. [[CrossRef](#)]
108. Asamitsu, K.; Yamaguchi, T.; Nakata, K.; Hibi, Y.; Victoriano, A.F.B.; Imai, K.; Onozaki, K.; Kitade, Y.; Okamoto, T. Inhibition of human immunodeficiency virus type 1 replication by blocking IkappaB Kinase with noraristeromycin. *J. Biochem.* **2008**, *144*, 581–589. [[CrossRef](#)]
109. Bhargava, P.; Malik, V.; Liu, Y.; Ryu, J.; Kaul, S.C.; Sundar, D.; Wadhwa, R. Molecular Insights Into Withaferin-A-Induced Senescence: Bioinformatics and Experimental Evidence to the Role of NF $\kappa$ B and CARF. *J. Gerontol. A Biol. Sci. Med. Sci.* **2019**, *74*, 183–191. [[CrossRef](#)]
110. Duan, X.; Ponomareva, L.; Veeranki, S.; Panchanathan, R.; Dickerson, E.; Choubey, D. Differential roles for the interferon-inducible IFI16 and AIM2 innate immune sensors for cytosolic DNA in cellular senescence of human fibroblasts. *Mol. Cancer Res.* **2011**, *9*, 589–602. [[CrossRef](#)]
111. Fafián-Labora, J.A.; O’Loughlen, A. NF- $\kappa$ B/IKK activation by small extracellular vesicles within the SASP. *Aging Cell* **2021**, *20*, e13426. [[CrossRef](#)] [[PubMed](#)]
112. Li, T.; Yan, B.; Xiao, X.; Zhou, L.; Zhang, J.; Yuan, Q.; Shan, L.; Wu, H.; Efferth, T. Onset of p53/NF- $\kappa$ B signaling crosstalk in human melanoma cells in response to anti-cancer theabrownin. *FASEB J.* **2022**, *36*, e22426. [[CrossRef](#)] [[PubMed](#)]
113. Choubey, D.; Panchanathan, R. IFI16, an amplifier of DNA-damage response: Role in cellular senescence and aging-associated inflammatory diseases. *Ageing Res. Rev.* **2016**, *28*, 27–36. [[CrossRef](#)] [[PubMed](#)]

114. Lorenz, J.; Zahlten, J.; Pollok, I.; Lippmann, J.; Scharf, S.; N'Guessan, P.D.; Opitz, B.; Flieger, A.; Suttorp, N.; Hippenstiel, S.; et al. Legionella pneumophila-induced I $\kappa$ B $\zeta$ -dependent expression of interleukin-6 in lung epithelium. *Eur. Respir. J.* **2011**, *37*, 648–657. [[CrossRef](#)] [[PubMed](#)]
115. Cuollo, L.; Antonangeli, F.; Santoni, A.; Soriani, A. The Senescence-Associated Secretory Phenotype (SASP) in the Challenging Future of Cancer Therapy and Age-Related Diseases. *Biology* **2020**, *9*, 485. [[CrossRef](#)] [[PubMed](#)]
116. Burton, D.G.; Faragher, R.G. Cellular senescence: From growth arrest to immunogenic conversion. *Age*. **2015**, *37*, 27. [[CrossRef](#)] [[PubMed](#)]
117. Moiseeva, O.; Deschenes-Simard, X.; St-Germain, E.; Igelmann, S.; Huot, G.; Cadar, A.E.; Bourdeau, V.; Pollak, M.N.; Ferbeyre, G. Metformin inhibits the senescence-associated secretory phenotype by interfering with IKK/NF-kappaB activation. *Aging Cell* **2013**, *12*, 489–498. [[CrossRef](#)]
118. Birch, J.; Gil, J. Senescence and the SASP: Many therapeutic avenues. *Genes Dev.* **2020**, *34*, 1565–1576, PMID: 33262144; PMCID: PMC7706700. [[CrossRef](#)]
119. Tilstra, J.S.; Robinson, A.R.; Wang, J.; Gregg, S.Q.; Clauson, C.L.; Reay, D.P.; Nasto, L.A.; St Croix, C.M.; Usas, A.; Vo, N.; et al. NF-kappaB inhibition delays DNA damage-induced senescence and aging in mice. *J. Clin. Investig.* **2012**, *122*, 2601–2612. [[CrossRef](#)]
120. Song, W.; Li, D.; Tao, L.; Luo, Q.; Chen, L. Solute carrier transporters: The metabolic gatekeepers of immune cells. *Acta Pharm. Sin. B* **2020**, *10*, 61–78. [[CrossRef](#)]
121. Heuser, C.; Gotot, J.; Piotrowski, E.C.; Philipp, M.S.; Courrèges, C.J.F.; Otte, M.S.; Guo, L.; Schmid-Burgk, J.L.; Hornung, V.; Heine, A.; et al. Prolonged IKK $\beta$  Inhibition Improves Ongoing CTL Antitumor Responses by Incapacitating Regulatory T Cells. *Cell Rep.* **2017**, *21*, 578–586. [[CrossRef](#)] [[PubMed](#)]
122. Senftleben, U.; Li, Z.W.; Baud, V.; Karin, M. IKKbeta is essential for protecting T cells from TNFalpha-induced apoptosis. *Immunity* **2001**, *14*, 217–230. [[CrossRef](#)] [[PubMed](#)]
123. Deng, H.; Mao, G.; Zhang, J.; Wang, Z.; Li, D. IKK antagonizes activation-induced cell death of CD4+ T cells in aged mice via inhibition of JNK activation. *Mol. Immunol.* **2010**, *48*, 287–293. [[CrossRef](#)] [[PubMed](#)]
124. Blonska, M.; Joo, D.; Nurieva, R.L.; Zhao, X.; Chiao, P.; Sun, S.C.; Dong, C.; Lin, X. Activation of the transcription factor c-Maf in T cells is dependent on the CARMA1-IKK $\beta$  signaling cascade. *Sci. Signal.* **2013**, *6*, ra110. [[CrossRef](#)] [[PubMed](#)]
125. Krishna, S.; Xie, D.; Gorentla, B.; Shin, J.; Gao, J.; Zhong, X.P. Chronic activation of the kinase IKK $\beta$  impairs T cell function and survival. *J. Immunol.* **2012**, *189*, 1209–1219. [[CrossRef](#)] [[PubMed](#)]
126. Miller, M.L.; Mashayekhi, M.; Chen, L.; Zhou, P.; Liu, X.; Michelotti, M.; Tramontini Gunn, N.; Powers, S.; Zhu, X.; Evaristo, C.; et al. Basal NF- $\kappa$ B controls IL-7 responsiveness of quiescent naïve T cells. *Proc. Natl. Acad. Sci. USA* **2014**, *111*, 7397–7402. [[CrossRef](#)] [[PubMed](#)]
127. Jeucken, K.C.M.; van Rooijen, C.C.N.; Kan, Y.Y.; Kocken, L.A.; Jongejan, A.; van Steen, A.C.I.; van Buul, J.D.; Olsson, H.K.; van Hamburg, J.P.; Tas, S.W. Differential Contribution of NF- $\kappa$ B Signaling Pathways to CD4+ Memory T Cell Induced Activation of Endothelial Cells. *Front. Immunol.* **2022**, *13*, 860327. [[CrossRef](#)]
128. Chuang, H.C.; Tsai, C.Y.; Hsueh, C.H.; Tan, T.H. GLK-IKK $\beta$  signaling induces dimerization and translocation of the AhR-ROR $\gamma$ t complex in IL-17A induction and autoimmune disease. *Sci. Adv.* **2018**, *4*, eaat5401. [[CrossRef](#)]
129. Evaristo, C.; Spranger, S.; Barnes, S.E.; Miller, M.L.; Molinero, L.L.; Locke, F.L.; Gajewski, T.F.; Alegre, M.L. Cutting Edge: Engineering Active IKK $\beta$  in T Cells Drives Tumor Rejection. *J. Immunol.* **2016**, *196*, 2933–2938. [[CrossRef](#)]
130. Ding, W.; Wagner, J.A.; Granstein, R.D. CGRP, PACAP, and VIP modulate Langerhans cell function by inhibiting NF-kappaB activation. *J. Investig. Dermatol.* **2007**, *127*, 2357–2367. [[CrossRef](#)]
131. Bosch, N.C.; Voll, R.E.; Voskens, C.J.; Gross, S.; Seliger, B.; Schuler, G.; Schaft, N.; Dörrie, J. NF- $\kappa$ B activation triggers NK-cell stimulation by monocyte-derived dendritic cells. *Ther. Adv. Med. Oncol.* **2019**, *11*, 1758835919891622. [[CrossRef](#)] [[PubMed](#)]
132. Yang, J.; Hawkins, O.E.; Barham, W.; Gilchuk, P.; Boothby, M.; Ayers, G.D.; Joyce, S.; Karin, M.; Yull, F.E.; Richmond, A. Myeloid IKK $\beta$  promotes antitumor immunity by modulating CCL11 and the innate immune response. *Cancer Res.* **2014**, *74*, 7274–7284. [[CrossRef](#)] [[PubMed](#)]
133. Lopez-Pelaez, M.; Lamont, D.J.; Pegg, M.; Shpiro, N.; Gray, N.S.; Cohen, P. Protein kinase IKK $\beta$ -catalyzed phosphorylation of IRF5 at Ser462 induces its dimerization and nuclear translocation in myeloid cells. *Proc. Natl. Acad. Sci. USA* **2014**, *111*, 17432–17437. [[CrossRef](#)] [[PubMed](#)]
134. Baratin, M.; Foray, C.; Demaria, O.; Habbeldine, M.; Pollet, E.; Maurizio, J.; Verthuy, C.; Davanture, S.; Azukizawa, H.; Flores-Langarica, A.; et al. Homeostatic NF- $\kappa$ B Signaling in Steady-State Migratory Dendritic Cells Regulates Immune Homeostasis and Tolerance. *Immunity* **2015**, *42*, 627–639. [[CrossRef](#)] [[PubMed](#)]
135. Wang, X.; Wang, J.; Zheng, H.; Xie, M.; Hopewell, E.L.; Albrecht, R.A.; Nogusa, S.; García-Sastre, A.; Balachandran, S.; Beg, A.A. Differential requirement for the IKK $\beta$ /NF- $\kappa$ B signaling module in regulating TLR- versus RLR-induced type 1 IFN expression in dendritic cells. *J. Immunol.* **2014**, *193*, 2538–2545. [[CrossRef](#)]
136. Chow, K.T.; Wilkins, C.; Narita, M.; Green, R.; Knoll, M.; Loo, Y.M.; Gale, M., Jr. Differential and Overlapping Immune Programs Regulated by IRF3 and IRF5 in Plasmacytoid Dendritic Cells. *J. Immunol.* **2018**, *201*, 3036–3050. [[CrossRef](#)]
137. Karin, M.; Ben-Neriah, Y. Phosphorylation meets ubiquitination: The control of NFkappaB activity. *Annu. Rev. Immunol.* **2000**, *18*, 621–663. [[CrossRef](#)]
138. Erol, A. IKK-mediated CYLD phosphorylation and cellular redox activity. *Mol. Med.* **2022**, *28*, 14. [[CrossRef](#)]

139. Pradère, J.P.; Hernandez, C.; Koppe, C.; Friedman, R.A.; Luedde, T.; Schwabe, R.F. Negative regulation of NF- $\kappa$ B p65 activity by serine 536 phosphorylation. *Sci. Signal.* **2016**, *9*, ra85. [[CrossRef](#)]
140. Zhang, J.; Zhao, R.; Yu, C.; Bryant, C.L.N.; Wu, K.; Liu, Z.; Ding, Y.; Zhao, Y.; Xue, B.; Pan, Z.Q.; et al. IKK-Mediated Regulation of the COP9 Signalosome via Phosphorylation of CSN5. *J. Proteome Res.* **2020**, *19*, 1119–1130. [[CrossRef](#)]
141. Blanchett, S.; Dondelinger, Y.; Barbarulo, A.; Bertrand, M.J.M.; Seddon, B. Phosphorylation of RIPK1 serine 25 mediates IKK dependent control of extrinsic cell death in T cells. *Front. Immunol.* **2022**, *1*, 1067164. [[CrossRef](#)] [[PubMed](#)]
142. Zhang, X.; Yu, H.; Zhao, J.; Li, X.; Li, J.; He, J.; Xia, Z.; Zhao, J. IKK $\epsilon$  negatively regulates RIG-I via direct phosphorylation. *J. Med. Virol.* **2016**, *88*, 712–718. [[CrossRef](#)] [[PubMed](#)]
143. Zhao, P.; Wong, K.I.; Sun, X.; Reilly, S.M.; Uhm, M.; Liao, Z.; Skorobogatko, Y.; Saltiel, A.R. TBK1 at the Crossroads of Inflammation and Energy Homeostasis in Adipose Tissue. *Cell* **2018**, *172*, 731–743.e12. [[CrossRef](#)] [[PubMed](#)]
144. Remoli, A.L.; Sgarbanti, M.; Perrotti, E.; Acchioni, M.; Orsatti, R.; Acchioni, C.; Battistini, A.; Clarke, R.; Marsili, G. I $\kappa$ B kinase- $\epsilon$ -mediated phosphorylation triggers IRF-1 degradation in breast cancer cells. *Neoplasia* **2020**, *22*, 459–469. [[CrossRef](#)] [[PubMed](#)]
145. Amaya, M.; Keck, F.; Bailey, C.; Narayanan, A. The role of the IKK complex in viral infections. *Pathog Dis.* **2014**, *72*, 32–44, Epub 2014 Aug 28. PMID: 25082354; PMCID: PMC7108545. [[CrossRef](#)] [[PubMed](#)]
146. Wang, V.Y.; Li, Y.; Kim, D.; Zhong, X.; Du, Q.; Ghassemian, M.; Ghosh, G. Bcl3 Phosphorylation by Akt, Erk2, and IKK Is Required for Its Transcriptional Activity. *Mol. Cell* **2017**, *67*, 484–497.e5. [[CrossRef](#)] [[PubMed](#)]
147. Ling, L.; Cao, Z.; Goeddel, D.V. NF- $\kappa$ B-inducing kinase activates IKK- $\alpha$  by phosphorylation of Ser-176. *Proc. Natl. Acad. Sci. USA* **1998**, *95*, 3792–3797. [[CrossRef](#)]
148. Antonia, R.J.; Baldwin, A.S. IKK promotes cytokine-induced and cancer-associated AMPK activity and attenuates phenformin-induced cell death in LKB1-deficient cells. *Sci. Signal.* **2018**, *11*, eaan5850. [[CrossRef](#)]
149. Lee, D.F.; Hung, M.C. Advances in targeting IKK and IKK-related kinases for cancer therapy. *Clin. Cancer Res.* **2008**, *14*, 5656–5662. [[CrossRef](#)]
150. Schomer-Miller, B.; Higashimoto, T.; Lee, Y.K.; Zandi, E. Regulation of I $\kappa$ B kinase (IKK) complex by IKK $\gamma$ -dependent phosphorylation of the T-loop and C terminus of IKK $\beta$ . *J. Biol. Chem.* **2006**, *281*, 15268–15276. [[CrossRef](#)]
151. Hehner, S.P.; Hofmann, T.G.; Ushmorov, A.; Dienz, O.; Wing-Lan Leung, I.; Lassam, N.; Scheidereit, C.; Dröge, W.; Schmitz, M.L. Mixed-lineage kinase 3 delivers CD3/CD28-derived signals into the I $\kappa$ B kinase complex. *Mol. Cell Biol.* **2000**, *20*, 2556–2568. [[CrossRef](#)] [[PubMed](#)]
152. Li, N.; Banin, S.; Ouyang, H.; Li, G.C.; Courtois, G.; Shiloh, Y.; Karin, M.; Rotman, G. ATM is required for I $\kappa$ B kinase (IKK) activation in response to DNA double strand breaks. *J. Biol. Chem.* **2001**, *276*, 8898–8903. [[CrossRef](#)] [[PubMed](#)]
153. Menden, H.; Tate, E.; Hogg, N.; Sampath, V. LPS-mediated endothelial activation in pulmonary endothelial cells: Role of Nox2-dependent IKK- $\beta$  phosphorylation. *Am. J. Physiol. Lung Cell. Mol. Physiol.* **2013**, *304*, L445–L455. [[CrossRef](#)]
154. Li, T.; Wong, V.K.; Jiang, Z.H.; Jiang, S.P.; Liu, Y.; Wang, T.Y.; Yao, X.J.; Su, X.H.; Yan, F.G.; Liu, J.; et al. Mutation of cysteine 46 in IKK- $\beta$  increases inflammatory responses. *Oncotarget* **2015**, *6*, 31805–31819. [[CrossRef](#)] [[PubMed](#)]
155. Nomura, F.; Kawai, T.; Nakanishi, K.; Akira, S. NF- $\kappa$ B activation through IKK-i-dependent I-TRAF/TANK phosphorylation. *Genes Cells* **2000**, *5*, 191–202. [[CrossRef](#)] [[PubMed](#)]
156. Motolani, A.; Martin, M.; Sun, M.; Lu, T. Phosphorylation of the Regulators, a Complex Facet of NF- $\kappa$ B Signaling in Cancer. *Biomolecules* **2020**, *11*, 15. [[CrossRef](#)] [[PubMed](#)]
157. Chariot, A. The NF- $\kappa$ B-independent functions of IKK subunits in immunity and cancer. *Trends Cell Biol.* **2009**, *19*, 404–413. [[CrossRef](#)] [[PubMed](#)]
158. Whitesid, S.T.; Ernst, M.K.; LeBail, O.; Laurent-Winter, C.; Rice, N.; Israël, A. N- and C-terminal sequences control degradation of MAD3/I $\kappa$ B $\alpha$  in response to inducers of NF- $\kappa$ B activity. *Mol. Cell Biol.* **1995**, *15*, 5339–5345. [[CrossRef](#)]
159. Palkowitsch, L.; Leidner, J.; Ghosh, S.; Marienfeld, R.B. Phosphorylation of serine 68 in the I $\kappa$ B kinase (IKK)-binding domain of NEMO interferes with the structure of the IKK complex and tumor necrosis factor- $\alpha$ -induced NF- $\kappa$ B activity. *J. Biol. Chem.* **2008**, *283*, 76–86. [[CrossRef](#)]
160. Zandi, E.; Rothwarf, D.M.; Delhase, M.; Hayakawa, M.; Karin, M. The I $\kappa$ B kinase complex (IKK) contains two kinase subunits, IKK $\alpha$  and IKK $\beta$ , necessary for I $\kappa$ B phosphorylation and NF- $\kappa$ B activation. *Cell* **1997**, *91*, 243–252. [[CrossRef](#)] [[PubMed](#)]
161. Garbati, M.R.; Gilmore, T.D. Ser484 and Ser494 in REL are the major sites of IKK phosphorylation in vitro: Evidence that IKK does not directly enhance GAL4-REL transactivation. *Gene Expr.* **2008**, *14*, 195–205. [[CrossRef](#)] [[PubMed](#)]
162. Hinz, M.; Scheidereit, C. The I $\kappa$ B kinase complex in NF- $\kappa$ B regulation and beyond. *EMBO Rep.* **2014**, *15*, 46–61. [[CrossRef](#)] [[PubMed](#)]
163. Chen, Z.J. Ubiquitination in signaling to and activation of IKK. *Immunol. Rev.* **2012**, *246*, 95–106. [[CrossRef](#)] [[PubMed](#)]
164. Napetschnig, J.; Wu, H. Molecular basis of NF- $\kappa$ B signaling. *Annu. Rev. Biophys.* **2013**, *42*, 443–468. [[CrossRef](#)] [[PubMed](#)]
165. Ben, J.; Jiang, B.; Wang, D.; Liu, Q.; Zhang, Y.; Qi, Y.; Tong, X.; Chen, L.; Liu, X.; Zhang, Y.; et al. Major vault protein suppresses obesity and atherosclerosis through inhibiting IKK-NF- $\kappa$ B signaling mediated inflammation. *Nat. Commun.* **2019**, *10*, 1801. [[CrossRef](#)] [[PubMed](#)]
166. Walsh, M.C.; Lee, J.; Choi, Y. Tumor necrosis factor receptor-associated factor 6 (TRAF6) regulation of development, function, and homeostasis of the immune system. *Immunol. Rev.* **2015**, *266*, 72–92. [[CrossRef](#)] [[PubMed](#)]

167. Kanarek, N.; Ben-Neriah, Y. Regulation of NF- $\kappa$ B by ubiquitination and degradation of the I $\kappa$ Bs. *Immunol. Rev.* **2012**, *246*, 77–94. [[CrossRef](#)]
168. Karim, Z.A.; Vemana, H.P.; Khasawneh, F.T. MALT1-ubiquitination triggers non-genomic NF- $\kappa$ B/IKK signaling upon platelet activation. *PLoS ONE* **2015**, *10*, e0119363. [[CrossRef](#)]
169. Vallabhapurapu, S.; Matsuzawa, A.; Zhang, W. Nonredundant and complementary functions of TRAF2 and TRAF3 in a ubiquitination cascade that activates NIK-dependent alternative NF- $\kappa$ B signaling. *Nat. Immunol.* **2008**, *9*, 1364–1370. [[CrossRef](#)]
170. Ji, Y.X.; Zhang, P.; Zhang, X.J.; Zhao, Y.C.; Deng, K.Q.; Jiang, X.; Wang, P.X.; Huang, Z.; Li, H. The ubiquitin E3 ligase TRAF6 exacerbates pathological cardiac hypertrophy via TAK1-dependent signalling. *Nat. Commun.* **2016**, *1*, 11267. [[CrossRef](#)]
171. Ohtake, F.; Saeki, Y.; Ishido, S.; Kanno, J.; Tanaka, K. The K48–K63 Branched Ubiquitin Chain Regulates NF- $\kappa$ B Signaling. *Mol. Cell* **2016**, *64*, 251–266. [[CrossRef](#)] [[PubMed](#)]
172. Tang, Y.; Tu, H.; Zhang, J.; Zhao, X.; Wang, Y.; Qin, J.; Lin, X. K63-linked ubiquitination regulates RIPK1 kinase activity to prevent cell death during embryogenesis and inflammation. *Nat. Commun.* **2019**, *10*, 4157. [[CrossRef](#)] [[PubMed](#)]
173. Tarantino, N.; Tinevez, J.Y.; Crowell, E.F.; Boisson, B.; Henriques, R.; Mhlanga, M.; Agou, F.; Israël, A.; Laplantine, E. TNF and IL-1 exhibit distinct ubiquitin requirements for inducing NEMO-IKK supramolecular structures. *J. Cell Biol.* **2014**, *204*, 231–245. [[CrossRef](#)] [[PubMed](#)]
174. Kai, X.; Chellappa, V.; Donado, C.; Reyon, D.; Sekigami, Y.; Ataca, D.; Louissaint, A.; Mattoo, H.; Joung, J.K.; Pillai, S. I $\kappa$ B kinase  $\beta$  (IKKB) mutations in lymphomas that constitutively activate canonical nuclear factor  $\kappa$ B (NF $\kappa$ B) signaling. *J. Biol. Chem.* **2014**, *289*, 26960–26972. [[CrossRef](#)] [[PubMed](#)]
175. Spina, V.; Rossi, D. NF- $\kappa$ B deregulation in splenic marginal zone lymphoma. *Semin. Cancer Biol.* **2016**, *39*, 61–67. [[CrossRef](#)]
176. Barwick, B.G.; Gupta, V.A.; Vertino, P.M.; Boise, L.H. Cell of Origin and Genetic Alterations in the Pathogenesis of Multiple Myeloma. *Front. Immunol.* **2019**, *21*, 1121. [[CrossRef](#)]
177. Sasaki, Y.; Calado, D.P.; Derudder, E.; Zhang, B.; Shimizu, Y.; Mackay, F.; Nishikawa, S.; Rajewsky, K.; Schmidt-Supprian, M. NIK overexpression amplifies, whereas ablation of its TRAF3-binding domain replaces BAFF:BAFF-R-mediated survival signals in B cells. *Proc. Natl. Acad. Sci. USA* **2008**, *105*, 10883–10888. [[CrossRef](#)]
178. Gatla, H.R.; Zou, Y.; Uddin, M.M.; Singha, B.; Bu, P.; Vancura, A.; Vancurova, I. Histone Deacetylase (HDAC) Inhibition Induces I $\kappa$ B Kinase (IKK)-dependent Interleukin-8/CXCL8 Expression in Ovarian Cancer Cells. *J. Biol. Chem.* **2017**, *292*, 5043–5054. [[CrossRef](#)]
179. Albenzi, B.C.; Mattson, M.P. Evidence for the involvement of TNF and NF-kappaB in hippocampal synaptic plasticity. *Synapse* **2000**, *35*, 151–159. [[CrossRef](#)]
180. Lubin, F.D.; Sweatt, J.D. The I $\kappa$ B kinase regulates chromatin structure during reconsolidation of conditioned fear memories. *Neuron* **2007**, *55*, 942–957. [[CrossRef](#)]
181. Saliminejad, K.; Khorram Khorshid, H.R.; Soleymani Fard, S.; Ghaffari, S.H. An overview of microRNAs: Biology, functions, therapeutics, and analysis methods. *J. Cell. Physiol.* **2019**, *234*, 5451–5465. [[CrossRef](#)] [[PubMed](#)]
182. Yang, Z.; Fang, S.; Di, Y.; Ying, W.; Tan, Y.; Gu, W. Modulation of NF- $\kappa$ B/miR-21/PTEN pathway sensitizes non-small cell lung cancer to cisplatin. *PLoS ONE* **2015**, *10*, e0121547. [[CrossRef](#)] [[PubMed](#)]

**Disclaimer/Publisher’s Note:** The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.